

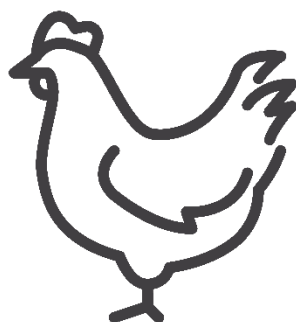


Pennsylvania
Department of Agriculture
Bureau of Animal Health and Diagnostic Services

CERTIFIED POULTRY TECHNICIAN

USER GUIDE

Revised April 4, 2025



Certified Poultry Technician (CPT) Course User Guide

KEY POINTS

***After completion of the CPT class, students must complete the field skills test within 6 months with the regional PDA veterinarian or inspector to demonstrate the techniques for sample collection, handling, and submission (refer to the CPT User Guide):**

- ♦ **Blood samples**
- ♦ **Tracheal, oropharyngeal, and cloacal swabs**
- ♦ **Rapid pullorum test (if applicable)**

***A skill checklist, signed by the classroom instructor, will be provided to you upon completion of the classroom training. Next steps:**

- a. **Schedule your field training with your regional office.**
- b. **Print the signed checklist.**
- c. **Provide the printed copy of the signed skills checklist to the field instructor for signature.**
- d. **Send the skills checklist with both instructors' signatures to the Department along with your license fee within 30 days after completion of the field skills test to obtain a license.**

***Licenses expire at the end of every year and must be renewed by January 1. Online renewal is available but not required.**

***Pennsylvania Certified Poultry Technicians are not authorized to collect samples from poultry outside of Pennsylvania.**

***CPTs collecting samples from PA poultry for entry into tested-source live bird markets must complete the LBM training and must request LBM CPT status before being added to the list of approved LBM CPTs.**

***It is recommended to contact the laboratory or check the PADLS website for a current testing fee schedule before sample submission.**

***A current email address is necessary for outreach and updates for all CPTs. If you don't have one, please provide one for someone who will share updates from PDA with you.**

Course Objectives

The Pennsylvania Department of Agriculture Certified Poultry Technician (CPT) Course is designed to provide trained technicians for collection of samples from poultry for regulatory testing purposes. This testing may be required for various reasons, including testing to meet poultry health program requirements, interstate movement, surveillance testing, or for show birds to enter exhibitions.

In addition, technicians will be trained to recognize symptoms of diseases in poultry, reporting protocols for suspected regulated diseases, and proper biosecurity practices to reduce the risk of spreading disease. Certified Poultry Technicians are vital to the protection of avian health through identifying and reporting birds suspected to be infected with dangerous transmissible diseases and through proper sample collection, handling, and submission.

The safety of technicians is paramount and can be promoted through learning proper sampling techniques and biosecurity practices.

At the completion of this course, the individual should have the knowledge and skills to:

- ♦ Recognize signs of avian disease
- ♦ Report suspected dangerous transmissible diseases to the Department
- ♦ Develop and follow proper biosecurity and cleaning and disinfection protocols
- ♦ Be familiar with proper personal protective equipment and personal safety
- ♦ Be familiar with the Department's avian health programs, including testing requirements
- ♦ Be familiar with the Department's requirements for testing for exhibition
- ♦ Be competent with blood and swab sample collection, handling, and submission
- ♦ If applicable, be competent performing and interpreting the Pullorum Rapid Whole Blood Agglutination Plate Test for breeding flocks
- ♦ Understand proper bird identification and record-keeping requirements (official leg bands)
- ♦ If sampling birds for entry into a live bird market, be competent in filling out Poultry Inspection Certificates (PICs) which accompany the birds and understand the rules for entry into the markets

The Pennsylvania Department of Agriculture may monitor any Certified Poultry Technician's performance, including sample collection, handling, and submission, as well as record keeping.

Please note: Please ensure we have a current email address for you. You can update your email address by emailing Stacy Etzweiler at setzweiler@pa.gov. You will be notified of important updates and events as needed via email. In addition, please include your email address on your license renewal application.

CERTIFIED POULTRY TECHNICIAN USER GUIDE

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SECTION I: CPT Skills Checklist (blank)



Pennsylvania
Department of Agriculture
Bureau of Animal Health and Diagnostic Services

2301 N. Cameron St. Harrisburg, PA 17110
Telephone: 717-783-6897 Fax: 717-787-1868

SKILL CHECKLIST FOR CERTIFIED POULTRY TECHNICIAN LICENSE

(Please provide to the field instructor after the classroom instructor has completed the classroom training section and signed the form).

CONTACT INFO:

NAME (LAST, FIRST):
ADDRESS:
EMAIL:

SKILLS COMPLETED SATISFACTORILY:

CLASSROOM: DATE COMPLETED:

- ☐ Understanding of proper sample collection, handling, and submission
☐ Understanding of proper bird identification and records retention (leg bands)
☐ Understanding of biosecurity
☐ Understanding of LBM requirements and Poultry Inspection Certificates
☐ Understanding of the rapid whole blood agglutination plate test (pullorum)

IN FIELD: DATE COMPLETED:

- ☐ Blood sample collection
☐ Tracheal/oropharyngeal swab collection
☐ Cloacal swab collection
☐ Rapid whole blood agglutination plate test (pullorum) if applicable

PRINTED NAME AND SIGNATURE OF CLASSROOM INSTRUCTOR:

(PRINTED NAME) (SIGNATURE-DO NOT PRINT) DATE

PRINTED NAME AND SIGNATURE OF FIELD INSTRUCTOR:

(PRINTED NAME) (SIGNATURE-DO NOT PRINT) DATE

Once completed, please return the form to the address below along with a \$10 check made out to the *Commonwealth of PA* for your license.

BAHDS POULTRY
2301 N. Cameron St. Harrisburg, PA 17110
Telephone: 717-783-6897 Fax: 717-787-1868

CPT Skills Checklist

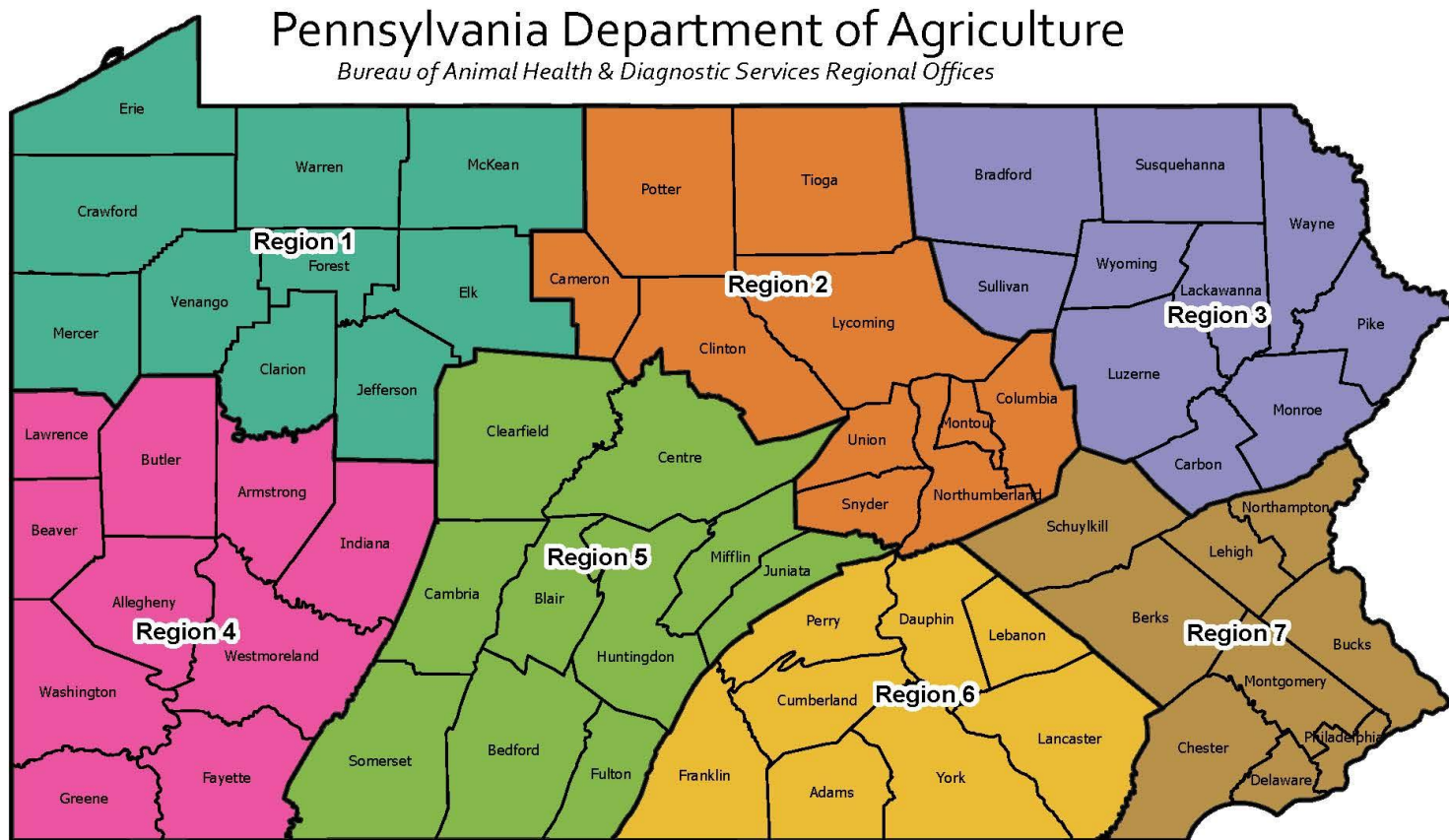
SECTION II: CONTACT INFORMATION AND REPORTING GUIDELINES

PA DEPARTMENT OF AGRICULTURE BUREAU OF ANIMAL HEALTH & DIAGNOSTIC SERVICES AND PENNSYLVANIA ANIMAL DIAGNOSTIC LABORATORY SYSTEM CONTACT LISTS

PA DEPARTMENT OF AGRICULTURE (PDA)		TELEPHONE	FAX
BUREAU OF ANIMAL HEALTH & DIAGNOSTIC SERVICES	2301 North Cameron Street, Harrisburg, PA 17110	717-772-2852 (24/7)	717-787-1868

PA ANIMAL DIAGNOSTIC LABORATORY SYSTEM (PADLS)		TELEPHONE	FAX
PA VETERINARY LABORATORY (PVL)	2305 North Cameron Street, Harrisburg, PA 17110	717-787-8808	717-772-3895
NEW BOLTON CENTER (NBC)	382 West Street Road Kennett Square, PA 19348 Avian Medicine and Pathology	610-444-4282	610-925-8106
PENN STATE UNIVERSITY Animal Diagnostic Laboratory (PSU ADL)	Animal Diagnostic Laboratory 131 Pastureview Road University Park, 16802	814-863-0837	814-865-3907

PA Department of Agriculture Regional Map



PA Department of Agriculture BAHDS 2301 N Cameron St Harrisburg PA 17110 (717) 772-2852 www.agriculture.pa.gov	Region 1 13410 Dunham Rd Meadville, PA 16335 (814) 332-6890 RA-AGAHR1@pa.gov	Region 2 542 County Farm Rd, Suite #102 Montoursville, PA 17754 (570) 433-2640 RA-AGAHR2@pa.gov	Region 3 No office address (570) 836-2181 RA-AGAHR3@pa.gov	Region 4 226 Donohoe Rd Greensburg, PA 15601 (724) 832-1073 RA-AGAHR4@pa.gov
	Region 5 (DEP District Office) 3001 Fairway Dr Altoona, PA 16602 (717) 705-5500 RA-AGAHR5@pa.gov	Region 6 2301 N Cameron St, Room 403 Harrisburg, PA 17110 (717) 346-3223 RA-AGAHR6@pa.gov	Region 7 1015 Bridge St, Suite #100 Collegeville, PA 19426 (610) 489-1003 RA-AGAHR7@pa.gov	

Guidelines for Reporting Dangerous Transmissible Diseases

- Report any signs suggestive of avian influenza (AI) or other dangerous transmissible diseases in a flock immediately to the Pennsylvania Department of Agriculture at: 717-772-2852, option 1 to reach the veterinarian on call. Do not transport suspects to a laboratory for diagnostic testing until AI or other dangerous transmissible diseases have been ruled out.
- Signs suggestive of AI or other reportable diseases may include (but are not limited to) the following:
 - Sudden increase in mortality
 - Decreased egg production
 - Decreased feed and water consumption
 - Swollen eyelids/sinuses/combs or wattles
 - Purple or bluish discoloration of wattles and combs
 - Respiratory signs
 - Generally depressed birds

SECTION III: COLLECTION, HANDLING, AND SUBMISSION OF SAMPLES

Key Points for Collection, Handling, and Submission of Samples

- ❖ Use appropriate personal protective equipment and follow biosecurity guidelines.
- ❖ Avoid handling poultry or samples if you are ill, especially if you are ill with influenza-like symptoms—in addition to the concern of zoonotic diseases, avian influenza flock test results could be affected.
- ❖ Blood and swab sample tubes, swabs, cardboard boxes for tubes, Whirl-pak® bags, viral transport media, and submission forms are available from PADLS laboratories. Needles and syringes are not provided by the PADLS laboratories.
- ❖ Cloacal swab samples are necessary for AI testing of waterfowl.
- ❖ Check the PADLS website for current user fees.
- ❖ Call the PADLS laboratory in advance to let them know when the samples will arrive if you are submitting a large number of samples or have specific testing requirements.
- ❖ In a disease outbreak situation, the PADLS website will be used to provide updated information and directions for sampling/testing related to disease control zones.
- ❖ Samples can be hand-carried to the laboratory (preferred method) or shipped by overnight mail or courier. Samples should be kept cool and delivered to the laboratory between Monday morning and Thursday noon. Samples should not be delivered on a day before a holiday or on weekends. If shipping samples, send early in the week, preferably before Wednesday morning. Do not use priority mail.
- ❖ Leg bands:
 - Birds tested for salmonella pullorum at the laboratories must be identified with an official PDA leg band and the band numbers must be recorded. Official leg bands are available from the PDA regional offices.
 - Each blood sample tube sent to the laboratory for pullorum testing must be labeled with the band number of the bird from which the sample was collected, or be numbered in such a way as to correspond to a band number written on the submission form, so that the sample can be traced back to an individual bird.
 - Records of leg band applications must be maintained for a minimum of two years and must be made available to PDA upon request.
 - If testing a breeder flock or exhibition flock on site using the rapid pullorum test, all reactors must be banded, and either blood from the reactor is sent immediately to the laboratory for further testing, or the bird is euthanized and sent immediately to the laboratory for tissue culture.
 - All birds entering a show must be banded with an official leg band, even if tested on site using the rapid pullorum test or if exempt from the pullorum testing requirement.
 - Official leg band applicators are not provided by PDA, but are available from the company listed below. Instead of using these applicators, you may use other tools to apply the leg bands.

National Band & Tag
Company 721 York St.,
PO Box 72430 Newport
KY 41072-0430 USA
859-261-2035
USA FAX: 800-261-8247
nationalband.com

Guidelines for Collection and Handling of Blood Samples

- Ensure you collect the correct number of samples on the correct schedule for the purpose of testing.
- **Collect samples from birds of age which are representative of the entire flock (all species, pens, and houses).**
- There are guidelines available for how much blood you can safely collect from a bird by weight. Please discuss this with your veterinarian.
- Only the clear serum, obtained after the blood sample has clotted and has been separated at the laboratory, is used for testing. You must submit enough blood from each bird to provide enough serum for all testing.
- If you are unable to safely collect enough blood for the necessary testing, consider swabbing for Avian Influenza (AI) testing.
- Usually, the brachial vein on the underside of the wing is used for blood collection.
 - Place the bird on its side, gently lift the wing and expose the vein, and part the feathers over the vein to visualize the vein.
 - Pull the plunger back slightly to break the vacuum of the seal.
 - Place the needle at a very slight angle to the vein, bevel up, insert the needle into the vein, and slowly withdraw the blood sample.
 - After collection, apply pressure to the vein for a short time as you withdraw the needle to reduce the risk of a hematoma forming.
 - To empty the syringe into the blood tube, use slow, gentle pressure on the plunger to avoid damaging the red blood cells.

Large tubes:

- Large (5 ml) tubes should be used when more than one test will be performed on the samples so a greater quantity of blood can be submitted.
- If you are using a box to hold and transport the blood tubes, place the necessary number of empty tubes in the cardboard tube box and, if possible, tilt the box approximately 30 degrees. Brace the box in this position while you collect samples to facilitate clot formation in the samples. If the temperature in the sampling area is high (summer months), hold the collected samples in a cooler with cold packs as you continue sampling. If using large, open-top tubes for hand delivery to the laboratory, instead of caps you can place a large piece of paper tape over the tops of the tubes in the box and keep the box upright to prevent leakage. Do not use open-top tubes with tape over the top for shipping – the labs will provide caps for the large tubes if requested for shipping.
- If you are using a Whirl-pak® bag to transport the blood tubes, cap the tubes tightly with caps provided by the laboratory and keep the tubes upright to prevent leakage.
- Collect approximately 2.5 milliliters (cc) of blood per bird and place it in a large (5 ml) tube.
- Complete the submission form and place it in a small plastic bag in the box on top of the tubes; or place it in the pocket of the Whirl-pak® bag which contains the blood samples.
- If the samples are to be tested for pullorum, either label the tubes to correspond to the official band number of each bird sampled and include the band number on the submission form, or label the tubes with the band number.
- If collecting samples from more than one bird type, either label the tubes to correspond to each bird type or place only one bird type per Whirl-pak® bag or box, and write the bird type on the corresponding bag or box (for example; chickens in one bag or box, turkeys in another).
- Place the box lid on tightly or seal the Whirl-pak® bag holding the tubes tightly.

- Label the box (not the lid) or Whirl-pak® bag with the owner name, premise ID or flock address, and date of collection.
- If using a box, place the box into a plastic bag, seal the bag, and disinfect the bag.
- If using a Whirl-pak® bag to hold the samples, disinfect the outside of the plastic bag.
- Place the samples in a cooler with frozen cold packs immediately after collection to keep them cool, and transport them in a cooler with frozen cold packs. Do not use ice and do not allow the blood samples to freeze.
- Submit samples as soon as possible after collection to ensure sample integrity.
- **Blood samples will not be accepted for testing of waterfowl for avian influenza. Cloacal swab testing is required for testing of waterfowl for avian influenza.**

Plastic snap cap tubes:

- If only one test will be performed on the samples, smaller plastic snap cap tubes, available from the PADLS laboratories, are acceptable.
- If using plastic snap cap tubes, collect 2.0 cc of blood per bird and place it in a tube. Fill the tube, but leave a small air space at the top so the lid does not pop open during transport.
- If the temperature in the sampling area is high (summer months), hold the collected samples in a cooler with cold packs as you continue sampling.
- Plastic snap cap tubes with blood samples should be placed into a labeled Whirl-pak® bag for submission to the laboratory. If collecting samples from more than one bird type, either label the tubes to correspond to each bird type or place only one bird type per Whirl-pak® bag, and write the bird type on the corresponding bag (for example; chickens in one bag, turkeys in another). Seal the bag(s).
- Label the Whirl-pak® bag with the owner name, premise ID or flock address, and date of collection.
- Complete the submission form and place it in the bag pocket.
- Disinfect the outside of the bag holding the samples.
- Place samples in a cooler with frozen cold packs immediately after collection to keep them cool, and transport them with frozen cold packs in a cooler. Do not use ice and do not allow the blood samples to freeze.
- Submit samples as soon as possible after collection to ensure sample integrity.
- **Blood samples will not be accepted for testing of waterfowl for avian influenza. Cloacal swab testing is required for avian influenza testing of waterfowl.**

Guidelines for Collection and Handling of Swab Samples for AI Testing

Tracheal and oropharyngeal (OP) swabs (for testing of gallinaceous birds such as chickens, turkeys, and game birds for AI):

- Tracheal swab samples are recommended for testing of freshly dead birds but may also be used on live birds. OP swab samples may be used for live bird sampling. Both methods include swabbing of the choanal slit in the roof of the mouth as you withdraw the swab.
- Ensure you collect the correct number of samples on the correct schedule for the purpose of testing.
- Swab testing may be used instead of blood testing to test gallinaceous birds for avian influenza. Swabs may not be used for pullorum testing of any type of bird.
- Brain Heart Infusion (BHI) broth or another liquid media will be provided for swab submissions. BHI should be clear. Do not use BHI that is cloudy or that has passed its expiration date.
- Keep BHI broth tubes refrigerated at all times.
- Use tubes containing 5.5 mls BHI.
- **Collect tracheal or oropharyngeal swabs for virus detection testing from birds of age which are representative of the entire flock.**
- Insert a dry sterile swab and rub the mucosa vigorously, dragging the swab through the choanal slit in the roof of the mouth as you finish swabbing.
- Use one swab for each bird.
- Avoid contaminating the swab through allowing contact with other surfaces.
- **Place material from up to 11 swabs into one BHI tube (gallinaceous birds only). Do not combine swab material from different bird types in a tube.**
- **If you are testing five birds or less of one bird type, you may leave up to five swabs in the tubes after breaking off the swab handles.**
- **If testing more than five birds (gallinaceous birds only), do not leave more than 5 swabs in the tubes, or remove all swabs** — after swabbing the bird, insert each swab into the tube, swirl the tip in the BHI, press the tip against the inner surface of the tube to remove excess material into the tube, and then discard the swab in a biosecure manner (maximum of 11 birds/tube).
- Secure the tube tops.
- Place the tubes into a Whirl-pak® bag or tube box. If you have sampled more than one bird type, either label the tubes to correspond to each bird type or place only one bird type per bag or box, and write the bird type on the corresponding bag or box (for example; chickens in one bag or box, turkeys in another). Close the bag or box securely.
- Label the bag or box with owner name, premise ID or address of the flock location, and date of collection.
- **Complete the submission form** and place it in the Whirl-pak® bag pocket or in a separate plastic bag inside the tube box.
- Disinfect the outside of the Whirl-pak® bag.
- If using a tube box, place the box inside of a gallon-sized plastic bag and disinfect the outside of the bag.
- Keep the samples cool on frozen cold packs in a cooler during transport. Do not use ice and do not allow the samples to freeze.
- Submit samples as soon as possible after collection to ensure sample integrity.
- Keep the BHI tubes in an upright position and ensure caps are tightly closed to prevent leakage. Tube boxes can be obtained from your PADLS laboratory.

Cloacal swabs (waterfowl) for AI testing (all waterfowl tested for AI must have cloacal swabs submitted for testing):

- Cloacal swab testing must be used to test waterfowl for avian influenza. Swabs may not be used for pullorum testing of any type of bird.
- Brain Heart Infusion (BHI) broth or another liquid media will be provided for swab submissions. BHI should be clear. Do not use BHI that is cloudy or that has passed its expiration date.
- Keep BHI broth tubes refrigerated at all times.
- Use tubes containing 5.5 mls BHI.
- **Collect cloacal swabs for virus detection testing from birds of age which are representative of the entire flock.**
- Insert the dry swab and rub the mucosa vigorously.
- Use one swab for each bird.
- Avoid contaminating the swab by allowing it to contact other surfaces.
- After swabbing, place swabs into tubes, ensuring that the ends of the swabs are covered completely by BHI broth.
- **Place up to 5 swabs (from up to 5 different birds of one type) into one tube. Leave the swabs in the tubes.**
- **Do not mix duck and goose swabs in a tube.**
- Break off the ends of the swab handles so the swabs fit into the tubes.
- Secure tube tops.
- Place tubes into a Whirl-pak® bag or box provided by the lab.
- If collecting samples from more than one type of waterfowl, either label the tubes to correspond to each bird type or place only one bird type per Whirl-pak® bag or tube box, and write the bird type on the corresponding bag or box.
- Label the bag or box with owner name, premise ID or flock address, and date of collection.
- **Complete the submission form** and place it in the Whirl-pak® bag pocket or in a separate plastic bag inside the tube box.
- Disinfect the outside of the Whirl-pak® bag.
- If using a tube box, place the box inside of a gallon-sized plastic bag and disinfect the outside of the bag.
- Keep the samples cool on frozen cold packs in a cooler during transport – do not use ice and do not allow the samples to freeze.
- Submit samples as soon as possible after collection to ensure sample integrity.
- Keep the BHI tubes in an upright position and ensure caps are tightly closed to prevent leakage. Tube boxes can be obtained from your PADLS laboratory.

Guidelines for Submission of Blood and Swab Samples

Hand Carried Blood or Swab Samples (Preferred Method)

After sample collection:

- If you are hand-carrying large blood tubes (5 ml) and are using a box to transport them, you may place paper tape over the top of the tubes in the tube box or use caps provided by the labs (caps are provided upon request).
- If you are using a Whirl-pak® bag to transport blood tubes, the tubes must be tightly capped, and the tubes placed into a labeled and sealed Whirl-pak® bag.
- Do not send blood samples in syringes and remove all needles from the boxes or bags. Packages with syringes and/or needles may be rejected.
- When submitting swab samples in BHI tubes, the tube caps must be tightly sealed, and the tubes placed in a tube box (preferred) or Whirl-pak® bag.
- All boxes and bags should be labelled with the owner name, flock address or premise ID, and date of collection.
- **All samples must be accompanied by a completed submission form.** Do not wrap the submission forms around samples (if a sample leaks, the form will be damaged). Enclose the paperwork for all samples in the outer pocket of the Whirl-pak® bag, or if using a box, in a zip-lock bag and placed on top of the samples in the box.
- Submit samples as soon as possible after collection, preferably within 24 - 48 hours, to ensure sample integrity. Samples received after 96 hours may be rejected.
- Keep the samples cool on frozen cold packs in a cooler during transport. Do not use ice and do not allow the samples to freeze.
- Deliver samples to the sample receiving area of the laboratory.
- Samples should be delivered to the laboratory between Monday morning and Thursday noon. Samples should not be delivered on a day before a holiday or on weekends. Hours of operation and holiday closings are available on the PADLS website: (padls.agriculture.pa.gov)

Guidelines for Submission of Blood and Swab Samples

Overnight Mail or Courier Delivery of Blood or Swab Samples

After collection:

- If you are shipping tubes, the tubes must be tightly capped. The labs will provide caps for the large blood tubes upon request.
- Do not send blood samples in syringes and remove all needles from the boxes or bags. Packages with syringes and/or needles may be rejected.
- When shipping swabs in BHI tubes, the tube caps must be tightly sealed, and the tubes placed upright in a tube box (preferred) or Whirl-pak® bag.
- All boxes and bags should be labelled with the owner name, flock address or premise ID, and date of collection.
- **All samples must be accompanied by a completed submission form.** Do not wrap the submission forms around samples (if a sample leaks, the form will be damaged). Enclose the paperwork for all samples in the outer pocket of the Whirl-pak® bag, or if using a box, in a zip-lock bag and placed on top of the samples in the box.
- Submit samples as soon as possible after collection, preferably within 24-48 hours, to ensure sample integrity. Samples received after 96 hours may be rejected.
- Keep the samples cool on frozen cold packs in a cooler during transport – do not use ice. Do not allow the samples to freeze.
- Include packing material in the container adequate to cushion the containers and absorb fluids in the event of leakage or breakage.
- If shipping samples, send early in the week, preferably before Wednesday morning. Samples should not be delivered on a day before a holiday or on weekends. Hours of operation and holiday closings are available on the PADLS website: (padls.agriculture.pa.gov)
[Welcome to PADLS](#)
- DO NOT use Priority Mail.
- Overnight trackable methods of shipment are recommended.
- For shipping, samples and submission forms should be packaged in a Styrofoam container with frozen cold packs and padding material. This container should be placed inside of a cardboard shipping box. Do not ship samples in unprotected Styrofoam containers because these can break easily if squeezed or dropped.
- Label the outside of the shipping box clearly. Remove old conflicting labels that could confuse delivery. Label the outside of the box “perishable”.
- Include your return address on the package.

Guidelines for Submission of Birds to the Laboratory for Diagnostic Testing

The following guidelines were designed to assist in the submission of fresh dead birds to the laboratory for diagnostic testing:

1. Submission Form: Complete the PADLS *General Submission Form* to accompany the birds.
2. History: Any health problems in the flock should be included on the submission form. A detailed description of any health problems in the flock, including changes in production, symptoms of illness, and mortality, should be included with the submission. In addition, field necropsy findings, recent feed changes, vaccinations, treatments, litter changes, etc. should be included to assist with the diagnosis.
3. Number of Birds in the Submission: As a general guideline, the following numbers may be used:
 - a. Growing and adult chickens and turkeys: 4 to 8 birds
 - b. Baby chicks and poults: 8 to 10 birds
4. Which Birds to Submit: It is important to choose birds for submission which will be most likely to provide the proper diagnosis of the problem in the flock. Please contact the diagnostician at the receiving lab to discuss the problem for further guidance on bird selection. The following guidelines apply:
 - a. If the flock has increased mortality, always include several fresh dead birds, representative of the mortality of the day.
 - b. If there are specific symptoms noted (respiratory signs such as snickering, foamy eyes, labored breathing; diarrhea or other changes in droppings; neurologic signs; leg problems; generally depressed; inactive; decreased feed consumption; failing to thrive; poor growth; weight loss; etc.), include live birds that are showing the specific symptom(s) of interest (call PDA or the laboratory before submission).
 - c. Include birds in different stages of illness if available.

Remember to contact PDA before submitting birds which are suspected of being infected with Avian Influenza or other dangerous transmissible diseases such as Newcastle Disease.

717-772-2852 option 1 (24/7)

Guidelines for Collection and Handling of Environmental Swabbing for NPIP Salmonella Programs

Below is a summary—refer to NPIP Program Standard B: *NPIP Procedures for collection, isolation, and identification of Salmonella*...These standards apply to egg and meat type chickens, turkeys, waterfowl, exhibition-type poultry, and game birds.

Information concerning the pen arrangement and number of birds per pen should be obtained from the owner so that the required number of samples per pen and per flock can be determined. A means of identifying each sample by pen of origin should be provided.

In addition to standard biosecurity procedures, the vehicle transporting the personnel taking the samples should be left as far as practical from the poultry pens. Biosecurity precautions, including using disinfected sampling equipment, sterile sampling supplies, and personal cleanliness, should be followed. Hands should be carefully washed with a sanitizing soap prior to the sampling. Outer clothing, including footwear and gloves, should be changed between visits to different premises so that clean clothing is worn upon entering each premises. A minimum size of three inches by three inches should be used for the sterile gauze pads.

After collection, the samples should be protected from drying, light, and excessive temperatures and delivered to the laboratory within one day. If delivery is delayed, samples should be refrigerated. Discuss this with the laboratory.

(1) Poultry House Environmental Samples. Fecal material, litter, dust, or nest box drag swab samples to be submitted for bacteriological examination shall be collected in accordance with the procedures described below:

(i) **Fecal material, litter or dust.** With a clean gloved hand or sterile collection device, collect fecal material, litter, or dust from several locations representing all areas of the pen or house into a sterile bag or container. A suggested minimum number of samples is five samples from pens or houses with less than 500 birds; 10 samples from pens or houses of 500 to 2,500 birds; and 15 samples from pens or houses with more than 2,500 birds.

(ii) **Drag swabs (DS).** DS, which consist of gauze pads or commercially available sponges, enable the sampling of large areas of the pen or house.

(A) Preparation. DS may be purchased commercially or be user prepared. One suggested method of making the DS assemblies is as follows: A sterile gauze pad is folded in half and a two-foot long (60cm) piece of twine is securely attached to the folded pad using a paper clip, staple, or similar device. A second sterile gauze pad is similarly fastened to a five-foot (150 cm) long piece of twine. The shorter piece of twine is then tied to the longer piece producing a DS sample set of two swabs arranged in a Y-shaped configuration. Alternatively, two separate DS samplers may be prepared. The twine is wrapped around the swabs, and the swabs *moistened* with double-strength skim milk (DSSM) (evaporated) or Buffered Peptone Water (BPW). The moistened swabs are placed in an instrument package. The sterilized swabs contained in the instrument package may be frozen (to prevent drying) until use.

(B) Procedure. At the farm the thawed DS assemblies are unraveled, and the ends of the twine held in gloved hands. The swabs are dragged across the environmental surfaces of the house for 15 minutes or the length of the house (down and back). One set of swabs (two individual pads) is dragged across the center of the house floor and another set of swabs (two individual pads) is dragged across the inside perimeter of the house floor. The four pads are individually placed in labeled, sterile bags. If necessary to prevent drying out, additional DSSM may be added to the bags. The bags should be protected from excessive heat and submitted as soon as possible to the authorized laboratory for testing. If the samples cannot be submitted to the laboratory the same day, they should be stored 2°- 8 °C or placed in a cooler with frozen cold packs for no more than five days before culturing.

(iii) **Shoe cover swabs (boot swabs).**

Absorbable fabric shoe covers involve the exposure of the bottom surface of shoe covers to the surface of floor litter and slat areas. Wearing clean gloves, place the shoe covers over footwear that is only worn inside the poultry house. This can be footwear dedicated to the facility or disposable overshoes. Each pair of shoe covers should be worn while walking at a normal pace over a distance of 1,000 feet (305 meters). For flocks with fewer than 500 breeders, at least one pair of shoe covers should be worn to sample the floor of the bird area. For flocks with 500 or more breeders, at least two pairs of shoe covers should be worn to sample the floor of the bird area. After sampling, place each shoe cover in a sterile container with 30 ml of DSSM, unless pre-moistened swabs (BPW) are used. Seal the sterile containers and promptly refrigerate them at 2° to 8 °C or place in a cooler with frozen cold packs. Do not freeze. If shoe cover swab samples are to be processed within 48 hours after collection, the shoe cover swab samples may be pre-moistened with BPW. Samples to be processed after 48 hours and before 5 days must be pre-moistened with DSSM. All samples are to be placed in a cooler with cold packs for transport and refrigeration at 2° - 8°C. Samples should be stored at refrigerator temperatures of 2°- 8°C no more than 5 days before culturing.

(iv) **Nest box or egg belt swabs as alternative sampling source.**

(A) Two sterile pre-moistened (ex. DSSM or BPW) gauze pads or sponges are swabbed along the inside of approximately 10 percent of the nest boxes. Each swab or sponge is placed into a separate sterile bag and submitted to the authorized laboratory.

(B) Two sterile pre-moistened (ex. DSSM or BPW) gauze pads or sponges are used to swab the egg belts. At least 30 feet of belt material is swabbed with each swab. Each swab is placed into a separate sterile bag and submitted to the authorized laboratory.

(C) If nest box or egg belt swab samples are to be processed within 48 hours after collection, the nest box or egg belt swab samples may be pre-moistened with BPW. Samples to be processed after 48 hours and before five days must be pre-moistened with DSSM. All samples are to be placed in a cooler with cold packs for transport and refrigeration at 2° - 8°C.

(2) **Hatchery Samples.** Hatchery-related samples, such as chick box papers, meconium, and fluff, may be examined for the presence of *Salmonella* to indicate the transfer of *Salmonella* from parent to offspring.

(i) **Chick box papers (swabs).** Chick box paper samples may be collected by an authorized agent (see below) or may be submitted directly to an authorized laboratory, with laboratory approval, for testing (see below). It is important to remove, with sanitized or gloved hands, the paper from the chick box before the box is placed in the brooding house.

(A) Instructions for sampling chick box papers. One chick box paper is collected for every 10 boxes of chicks placed in a house. With sanitized and gloved hands lay out the papers on a clean, disinfected surface. Saturate a sterile gauze pad or sponge with DSSM or BPW and swab the surface of five chick box papers. The pad should be rubbed over approximately 75 percent of each paper with sufficient pressure to remove any dried meconium. Addition of more DSSM or BPW may facilitate sampling. The process is repeated with a second swab and the other five chick box papers. Both swabs may be added to a single sterile, labeled plastic bag and submitted to the authorized laboratory. If chick box paper samples are to be processed within 48 hours after collection, the chick box paper samples may be pre-moistened with BPW. Samples to be processed after 48 hours and before five days must be pre-moistened with DSSM. All samples are to be placed in a cooler with cold packs for transport and refrigeration at 2° - 8°C. Promptly refrigerate the Whirl-pak® bags containing the samples and transport them with cold packs to a laboratory to be cultured within 5 days of collection.

(ii) **Chick box papers.** Chick box papers may be submitted directly to a laboratory. To send chick box papers directly to a laboratory (check with the laboratory before submission):

(A) With sanitized or gloved hands, collect one chick box paper for each 10 boxes of chicks placed in a house and place the chick papers immediately into large, clean plastic bags, and label and seal the bags.

(B) Place the plastic bags containing the chick box papers in a clean box and transport them within 48 hours to a laboratory. The plastic bags do not require refrigeration.

(iii) **Chick meconium.** After collection, the container of meconium is mixed to obtain a uniform consistency. In the laboratory a 25-gram sample will be removed for bacteriological examination.

(iv) **Fluff.** Fluff samples may be collected from the floor of the hatchery or from the tray following hatching. The fluff sample may be collected with sanitized or gloved hands by either swabbing the floor or tray with a pre-moistened gauze pad or sponge or by placing fluff material directly into a sterile bag.

PEQAP Guidelines for Environmental Swab Handling for Salmonella Testing

(Guidelines taken from PA Egg Quality Assurance Program operations annex)

Equipment

1. Standard biosecurity equipment
2. Small cooler with three frozen cold packs
3. Large garbage bag to serve as a tablecloth
4. Scissors
5. Can opener
6. Waterproof permanent marker
7. DSSM (double strength skim milk)
8. Optional: Manure drag pole (recommendation of constructing one from a 3/8" by 42" solid aluminum rod with a 1/4" hole drilled 1/2" from one end, or from a 1/2" by 36" conduit with a 1/4" hole drilled 1/2" from one end - the solid aluminum rods are easier to clean and disinfect).
9. PEQAP drag swab kit from Penn State Animal Diagnostic Laboratory or New Bolton Center; kit contains:
 - a) Whirl-pak® bags (18-ounce size)
 - b) Sterile gauze pads
10. Alcohol swabs

Manure Swabbing:

1. Prior to swabbing, label the sample bags with flock name and collection date.
2. Suit up with protective clothing and disinfect boots and swabbing equipment before entering the house in accordance with standard biosecurity practices.
3. Bring all materials to the bottom floor of the house. Use the bottom utility area if the house has one. Bring a bucket filled with a disinfecting solution.
4. Spread out the large garbage bag and arrange the sampling materials on top of the bag. Label the bags.
5. Put on a pair of disposable gloves.
6. Open the alcohol swab and wipe the top of the can of DSSM and the can opener. Wipe excess alcohol from the can and can opener with a clean paper towel before opening the can to minimize contamination of the milk with alcohol.
7. Disinfect the scissors with the disinfecting solution in your bucket. Wipe excess disinfectant off the scissors with a clean paper towel.
8. Use disinfected scissors to cut open the autoclave pack of swabs near the top of the pack.
9. Shake the can of DSSM and moisten the swabs in the pack by pouring a small amount of DSSM into the pack and massaging the outside of the pack. Lay the pack on the garbage bag. Clean and sanitize gloved hands after touching non-sterile items and before taking samples with sterile swabs. Wipe excess disinfectant from gloved hands with a clean paper towel.
10. Prepare the Whirl-pak® bags.
11. Tie two swabs to the strings per NPIP protocol.
12. Sample the manure per NPIP protocol.
13. Place the 2 swabs into separate Whirl-pak® bags *without touching the swabs* (one swab per bag). Cut attaching strings with the scissors and disinfect the scissors in between each sample; wipe excess disinfectant from the scissors with a clean paper towel or pull the strings off per NPIP guidelines.

Note- Adding too much DSSM can affect test results.
14. After the manure is dragged, place the Whirl-pak® bags (with samples) into the gallon- size bag, seal tightly, and place in the cooler with the frozen cold packs.

15. Put all discarded material into the garbage bag and dispose of properly.
16. Place the cooler outside the house; clean and disinfect it; then load it into your vehicle.
17. Follow standard biosecurity procedures when leaving.
18. Transport samples to the laboratory within 24-48 hours.

Egg Belts and Nest boxes:

Hand-swab the egg belts and nest boxes per NPIP or PEQAP protocol and handle as per manure swabbing guidelines.

SECTION IV: PADLS AVIAN SAMPLE SUBMISSION FORMS

AVIAN SAMPLE SUBMISSION FORM

See the PADLS website for the current version under the “tests and submissions”, “find forms” tab

<http://padls.agriculture.pa.gov>

Avian Sample Submission Form



Pennsylvania Animal Diagnostic Laboratory System Avian Sample Submission Form

University of Pennsylvania
New Bolton Center
382 West Street Road
Kennett Square, PA 19348
(610) 925-6725

Pennsylvania State University
Animal Diagnostic Laboratory
131 Pastureview Rd
University Park, PA 16802
(814) 863-0837

Pennsylvania Department of
Agriculture
Pennsylvania Veterinary
Laboratory
2305 North Cameron Street
Harrisburg, PA 17110-9408
(717) 787-8808

Billing and Reporting Preferences

Report to by:
Bill to: ☐ Fax: ☐ Email: ☐ US Mail: ☐
Sample Collector ☐ ☐ ☐ ☐
Owner/Company ☐ ☐ ☐ ☐
Premise Owner ☐ ☐ ☐ ☐

Accession

Sample Collector

Certified Poultry Tech ID Number _____

Name _____

Address _____

City, State, Zip _____

Phone _____ Fax _____

Email _____

Signature _____

Owner/Company

Owner _____

Company _____

Address _____

City, State, Zip _____

Phone _____ Fax _____

Email _____

See back of form if submitting multiple premises

MF# _____ Premises _____ NPIP# _____

Location of Birds at Sampling

Premises Identification Number _____

Flock ID/Name/House #/Floor #/Pen # _____

Address _____

City, State, Zip _____

Phone _____ Fax _____

Email _____

For a report sent to other than above. Name: _____ Fax/E-mail: _____

Pullet House Name: _____ Layer House Address: _____

Layer House Name: _____

Date Collected: _____ **Date Submitted:** _____ Age of flock: _____ Years _____ Weeks _____ Days

Blood: _____ # Eggs: _____ # Swabs: _____ Swab Source: _____

☐ Chicken ☐ Duck ☐ Guinea ☐ Turkey ☐ Other: _____ Breed: _____ Production type: _____

Description (color / distinctive markings) _____
(If submitting multiple species, flocks, or sample types, see back of form to identify samples)

Number of Birds on Premises: _____ Comments/History: _____

Hatchery name where birds originated: _____ If Breeders, hatchery name to incubate eggs: _____

PROGRAM TESTING (Purpose of test): (Check all that apply for this submission) – If applicable, enter individual bird/flock IDs on back.

• LBM (AI) - For PDA/USDA Use Only

☐ Auction/Swap Meet/Small Sale ☐ Backyard ☐ Dealer
☐ Feed Store ☐ Hauler ☐ Live Bird Market (At Market)
☐ Truck/Crate Wash

• Live Bird Market System (Avian Influenza)

☐ Production Unit (On Farm) – Moving to state of _____

• ☐ Export/Movement to:

• ☐ National Poultry Improvement Plan (NPIP)

☐ US AI Clean (Breeders) ☐ Subpart E
☐ US H5/H7 LPAI Monitored: (Non-Breeders)
☐ US MG Clean: ☐ US MS Clean: ☐ US MM Clean:
☐ Routine Test ☐ Suspect Retest
☐ US Pullorum-Typhoid Clean:
☐ Routine Test ☐ Reactor Retest ☐ Bird Culture
☐ US Salmonella Monitored ☐ US Sanitation Monitored
☐ US SE Clean:
☐ SE Monitored ☐ Routine Test ☐ Bird Culture

Related Accession number for retests: _____

☐ Diagnostic Test Requests: Enter the number of each type of test requested. (If applicable, enter individual bird IDs on back)

If chicken ELISA testing is requested, please indicate a preference of ELISA test system: ☐ IDEXX (ADL) ☐ BioChek (NBC)

_____ MG Plate	_____ NDV ELISA	_____ MG ELISA	_____ Pullorum –Typhoid Plate	_____ AI Virus Isolation
_____ MS Plate	_____ IBV ELISA	_____ MS ELISA	_____ Pullorum –Typhoid Tube	_____ AI RRT-PCR
_____ MM Plate	_____ IBD ELISA	_____ MG/MS ELISA	_____ Aerobic Culture	_____ MG PCR
_____ AI AGID	_____ REO ELISA	_____ HEV ELISA	_____ Salmonella Culture	_____ MS PCR
_____ Other _____	_____ AE ELISA	_____ BA ELISA	_____ SE Culture Only	
	_____ SE PCR Only	_____ RapidChek SE Test Only (NBC)		

Please use the avian necropsy submission form if for diagnostic necropsy/analysis on birds or tissues

Blood Tube Identification*

Box # _____ Pen/House # _____ Species _____

Box # _____ Pen/House # _____ Species _____

*Please write band number **or** sample # in the space corresponding to sample location in box.**Multiple Flock Submission Information/ Swab Identification (only one accession will be created per form)**

Sample source: T-Tracheal Swab, O-Oropharyngeal Swab, C-Cloacal Swab, E-Environmental Swab, Blood, Eggs, Birds

Date Collected	Tube # / Lot #	Species/Breed	Sample Source (Include # of samples)	Flock ID, Description and/or Comments	Age	Location Number

PADLS reserves the right to perform tests for any of the diseases regulated or under surveillance by the Pennsylvania Department of Agriculture on any specimen it receives. PADLS reserves the right to perform any test on animals submitted for autopsy that the Case Coordinator deems necessary for obtaining a diagnosis. Your submission of specimens for diagnostic purposes constitutes your acknowledgment that some tests may be performed at other laboratories.

HPAI SAMPLE SUBMISSION FORM

See the PADLS website for the current version under the “tests and submissions”, “find forms” tab

<http://padls.agriculture.pa.gov>

**ONLY TO BE USED AT DEPARTMENT
DIRECTION FOR SAMPLES RELATED TO
HPAI TESTING**

HPAI PCR Submission Form



PENNSYLVANIA ANIMAL DIAGNOSTIC LABORATORY SYSTEM

High Path Avian Influenza PCR Submission Form

Pennsylvania State University
Animal Diagnostic Laboratory
131 Pastureview Rd
University Park, PA 16802
814-863-0837
ADLSubmissions@psu.edu

New Bolton Center
Veterinary Laboratory
382 West Street Rd.
Kennett Square, PA 19348
610-925-6725
NBCHPAI@vet.upenn.edu

PA Veterinary Laboratory
PA Department of Agriculture
2305 N. Cameron St.
Harrisburg, PA 17110
717-787-8808
PVLSUBMIT@pa.gov

Billing and Reporting Preferences

Report to by:

	Bill to:	Fax:	Email:	US Mail:
Sample Collector	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Owner/Company	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Premises Owner	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Accession # _____

Submitter (Collector)	Owner/Company	MF#	Premises	NPIP#
<input type="checkbox"/> I am a Certified Poultry Technician <input type="checkbox"/> I am a HPAI Sampler Certified Poultry Tech ID Number _____ Name _____ Address _____ City, State, Zip _____ Phone _____ Fax _____ Email _____ Signature _____	Owner _____ Company _____ Address _____ City, State, Zip _____ Phone _____ Fax _____ Email _____	_____ Premises ID Number _____ Flock ID/Name/House #/Floor #/Pen # _____ Address _____ City, State, Zip _____ Phone _____ Fax _____ Email _____		

For a report sent to Name: _____
other than above: Fax/E-mail: _____

Date Collected: _____ **Date Submitted:** _____ **Age of flock:** _____ Years _____ Weeks _____ Days

Swabs: _____ **# Swab pools:** _____ **Swab source:** _____

☐ Chicken ☐ Duck ☐ Guinea ☐ Turkey ☐ Other: _____ **Breed:** _____ **Production type:** _____

***If submitting swab pools from multiple species or flocks, please use page 2 to identify samples.**

Description (color / distinctive markings): _____

Number of Birds on Premises: _____ **Comments/History:** _____

PROGRAM TESTING (Purpose of test): (Check all that apply for this submission)

HPAI Surveillance

☐ Infected Zone (0-3K)
☐ Required Weekly Test

☐ Buffer Zone (3-10K)
☐ Required Weekly Test

☐ Surveillance Zone (10-20k)
☐ Required Weekly Test

Regulatory Investigation

☐ Epidemiology Linked
☐ Sick Bird Call
☐ FADI # _____

Live Bird Market System (Avian Influenza)

☐ Production Unit (On Farm) – Moving to State of _____

☐ Pennsylvania AI Monitored Flock Program

National Poultry Improvement Plan (NPIP)

☐ US AI Clean (Breeders)
☐ US H5/H7 LPAI Monitored: (Non-Breeders)

☐ **Export/Movement To: (State)** _____
Date of Movement & Time: _____

Multiple Flock Submission Information & Swab Identification

Sample Source: T-Tracheal Swab, O-Oropharyngeal Swab, C-Cloacal Swab, E-Environmental Swab

Sample Bar Code or Tube #	Age	# of Swabs	Swab Source (T, O, C)	Species/Breed	House Flock ID	Production Type

SECTION V: GUIDELINES FOR CONDUCTING THE PULLORUM RAPID WHOLE BLOOD AGGLUTINATION PLATE TEST

Instructions for Conducting Pullorum Rapid Whole Blood Agglutination Plate Test

Refer to NPIP Program Standard A
(poultryimprovement.org)

Indications: Pullorum disease is caused by bacteria known as *Salmonella pullorum*. This bacterial infection endures in the ovaries of the laying female. The disease is transmitted from mother to chick through the incubated egg. If the egg hatches, the chick is already infected when it leaves the shell. Through the droppings other chicks may become quickly infected, until the disease spreads through the brood. The one and only effective control measure for *Pullorum* disease is the elimination of infected breeders. This involves blood testing, so that *Pullorum* “carriers” may be detected and removed from the breeding flocks.

Description: The serum plate agglutination (SPA) test is used to detect specific antibodies that will bind to an antigen and cause visible “clumping” or agglutination. The antigen is prepared from cultures of *Salmonella pullorum* and a dye is added to improve visibility of the reactions. A prescribed amount of antigen is placed on a solid support, such as a glass plate or mirror, keeping each drop of antigen separate. An equal amount of test serum is placed next to the antigen and these are then mixed together. After a short incubation, the mixture is examined for evidence of agglutination, which appears as discrete clumps of the stained particles with a clearer background. If no antibodies are detected, the mixture will remain opaque. This is a screening test only and additional testing must be performed on all reactors. **This test is not approved for use in turkeys.**

Who can perform this test in PA?

- CPTs must demonstrate proficiency before being approved to perform the test. Interpretation of the results is subjective and requires training and skill.
- Antigen must be used before its expiration date.

Equipment necessary for testing:

- Required Test Components: Pullorum Plate Antigen; positive and negative control sera; test samples (never frozen); a glass plate or mirror marked in squares of ~1-1.5 in; bleeding needle, blood loop (standardized), method of disinfecting, rinsing, and drying loop and needle, and a device to hold birds individually while waiting for the test results.
- Official leg bands, available from your PDA regional office.
- For NPIP birds: NPIP form 9-2 (available from PDA (717-783-6897)).
- For birds not on NPIP, an equivalent PDA rapid plate test form is available for exhibition bird testing (available from PDA at 717-783-6897).

Overview of the test procedure (follow specific instructions from the manufacturer):

1. The antigen must be used according to the directions of the test kit manufacturer. Ensure the antigen has not passed the expiration date.
2. Shake the antigen well.
3. It is advisable to test known positive and negative control sera before testing bird samples. Use separate pipettes for administering all controls and samples.
4. To conduct the testing of samples:
 - a. A drop of antigen is placed on the testing plate.
 - b. A loopful of blood is taken up from the wing vein after the vein has been lanced. When submerged in the blood and then carefully withdrawn, the loop becomes properly filled.
 - c. The loopful of blood is stirred into the drop of antigen, and the mixture spread to a diameter of about 1 inch.
 - d. The test plate is rocked from side to side a few times to mix the antigen and blood thoroughly, and to facilitate agglutination. Be sure to keep each mixture within a single square.
 - e. **The bird being tested must be kept isolated from the flock while the result is pending in case the sample has a positive reaction.**
5. Incubate at room temperature for 2 minutes or per label directions.
6. The reaction is read at the end of the incubation period. A positive reaction shows formation of discrete clumps of stained material, normally starting at the edge of the mixture. Negative reactions show little to no change in the opaque serum-antigen mixture.
7. The loop is rinsed in clean water and dried by touching it to a piece of clean blotting paper to avoid contamination of the next sample.
8. Send a completed NPIP 9-2 form or PDA equivalent form (for non-NPIP flocks) to PDA to record flock testing in compliance with NPIP and PDA requirements.

Reading the test:

- Positive reactions are indicated by a clumping of the antigen in well-developed, bluish-colored clusters surrounded by clear spaces, within 2 minutes. The greater the agglutination ability of the blood, the more rapid the clumping and the larger the clumps. A lesser reaction shows small, but clearly visible clumps surrounded by spaces only partially clear.
- Reactions which occur after 2 minutes should not be considered positive.
- Negative reactions show little to no change in the opaque serum antigen mixture.
- Biological reactions may vary between clear-cut positive and negative.

Precautions:

- Avoid exposing hands, eyes or clothing to the antigen. It is recommended that the test be conducted out of direct sunlight, where there is a minimum amount of dust. For product or technical information, please contact the manufacturer.
- Follow all label directions for storage and handling.

Things to avoid:

- Deteriorated antigen may give false readings. Before testing, check a drop of antigen, without blood, on the plate to check for spontaneous agglutination.
- Do not use antigen which is past the expiration date.
- Excessive evaporation, high temperatures, or incorrectly interpreting late flocculation may also lead to false readings.
- Testers should use care, not speed, for the number of birds tested is less important than maximum accuracy.

Cleaning the testing surface:

- Clean the plate with clear, warm or cool water. Hot water may coagulate blood, making it difficult to remove.
- Soaps, disinfectants, or cleaning compounds may leave a residue which may affect subsequent tests. Grease on plate may prevent blood antigen mixture from spreading properly, and grease may be removed with soap, after which plate must be thoroughly rinsed.
- After cleaning, polish plate with clean cloth, leaving no blood or lint on the surface.

References:

1. NPIP Program Standard A (2024)

Key Points:

***All birds tested individually for entry into a PA show must be banded at the time of test with an official PDA leg band, even if tested with the rapid plate test.**

***All birds with positive reactions on the plate test must be banded with an official PDA leg band. and must either be sent immediately to the laboratory for culture or must have a blood sample collected immediately and sent to the laboratory for testing. All birds tested with the rapid test must remain isolated from the flock while a blood sample result is pending.**

SECTION VI: ESSENTIAL EQUIPMENT

Essential Equipment List

It is recommended that you take only essential items onto a premises for biosecurity reasons.

- ☐ Cooler and frozen cold packs (do not take into the poultry house-in extremely hot temperatures, take a second cooler and cold pack into the poultry house to keep samples cool during collection).
- ☐ Personal Protective Equipment
 - Clean cloth coveralls or new disposable coveralls
 - Clean rubber boots or disposable booties
 - Disposable hair cover or washable hat
 - Disposable gloves (optional)
 - Disposable face masks (optional)
- ☐ Bucket and brush for cleaning boots
- ☐ Disinfectant and disinfecting wipes for small items
- ☐ Hand sanitizer or soap
- ☐ Plastic garbage bags for disposable items, also for holding dirty coveralls, etc
- ☐ Sample collection supplies as needed
 - Syringes/needles
 - Blood tubes
 - Cardboard tube box (if using large tubes for blood collection) and plastic gallon bags; or Whirl-pak® bags
 - Marking pen for labelling tubes
 - Paper tape if using large blood tubes in boxes and hand delivery
 - Sterile swab packs and viral transport media tubes if swabbing for AI
 - If not using a fresh needle for each bird (it is recommended that you use a fresh needle for each bird), saline and jar for cleaning needles between birds
 - Official PDA leg bands and applicators - Official leg bands must be obtained from your PDA regional office for show birds and/or birds tested for pullorum
 - Submission form and pen (do not take into the poultry house)
- ☐ Pullorum Rapid Whole Blood Agglutination Plate Test supplies as needed
 - Antigen
 - Testing Kit (heating source, testing surface)
 - Loop and lance
 - Official leg bands and applicators
 - Cleaning supplies
- ☐ Official Leg Bands
 - All poultry entering PA exhibitions must be leg banded with an official PDA leg band.
 - All birds tested at the laboratory for pullorum must be banded with an official PDA leg band. Each blood sample tube submitted to PADLS labs for pullorum testing must be labeled with the band number of the bird from which the sample was collected or a corresponding number so that the sample can be traced back to that bird.
 - If poultry are tested on site using the rapid pullorum test, all suspects (reactors) must be banded with official PDA leg bands and either blood from the reactor is sent to the lab for further testing, or the bird is sent to the lab for tissue culture.

SECTION VII: BIOSECURITY

Biosecurity Guidelines for CPTs on a Poultry Premises

Overview:

- Certain poultry diseases, including avian influenza, can be easily spread between flocks by people, clothes, vehicles, and equipment. The following are guidelines designed to reduce the risk of spreading disease between flocks.
- If visiting backyard premises, it is recommended that you visit only one premises per day. If you must visit more than one premises in a day, for each visit wear clean outerwear, including boots, and adhere to your best biosecurity management practices.
- Never allow congregation of poultry from more than one premises on one site for sampling. This is a biosecurity risk. Also, as a CPT you are required to observe the health of the entire flock, and sample birds that are representative of the entire flock.
- If visiting commercial premises, adhere to the biosecurity requirements of each premises. Most commercial premises will have a required down time of two or three days between premises unless these commercial flocks are under the same management and the management allows more than one site visit per day.

When entering a poultry premises:

- Park as far away from the poultry house/yard as possible.
- Avoid parking on an area where litter has been spread.
- Avoid stepping out onto the ground without boots designated to the premises—put on rubber boots which have been cleaned and disinfected or disposable booties.
- Put on clean cloth coveralls or new disposable coveralls.
- Put on a disposable hair cap or hat which can be cleaned and disinfected.
- Sanitize hands (disposable gloves are recommended).
- Face shields, eyewear, disposable face masks, or respirators are optional.
- Take only essential equipment and supplies with you. Equipment must always be carefully cleaned and disinfected prior to use on a poultry premises. This includes cell phones.

Before leaving a poultry premises:

- Do not enter your vehicle until you have removed your dirty outerwear. However, to prevent contamination of your vehicle, it is recommended to remove your outer boots after you are sitting on the vehicle seat, with your feet outside the vehicle. Wearing disposable booties under rubber boots is helpful as booties can be easily removed while sitting in the vehicle.
- Remove and bag all items worn on the premises.
- Dispose of any disposable items in a plastic garbage bag, close the bag tightly, and leave it on the premises for disposal if you have permission.
- Place any non-disposable clothing, such as cloth coveralls and raincoats, etc, worn on the premises in a clean garbage bag and wash the clothing in hot water before wearing it again.
- Clean and disinfect rubber boots before removal from the premises. Allow your boots to dry before you wear them again.
- If you have parked near the poultry or manure areas, and/or there is known disease in the flock, scrub vehicle tires with a brush to remove organic material and spray the tires with disinfectant before leaving the premises, and wash your vehicle before entering another poultry premises.
- Disinfect the outside of sample bags, coolers, and other non-disposable equipment and supplies before placing in your vehicle. This includes cell phones.
- Before placing your feet on the vehicle floorboards, spray the bottom of your footwear with disinfectant.
- Wash or sanitize your hands, including under fingernails.

Biosecurity Programs and Plans for Poultry Premises

In the aftermath of the HPAI outbreak in 2014/2015, and the HPAI outbreak which began in 2022, investigators with USDA reported that although wild birds introduced AI initially, poor biosecurity practices may have contributed to further spread.

Per USDA, any commercial premises (as defined by code of federal regulations) located in a HPAI control area around an infected flock is required to have an approved biosecurity plan, be following that plan, and pass a biosecurity audit of the premises to restock while in that control area to be eligible for indemnity. This also applies to commercial premises regardless of location, if previously infected with HPAI during the current outbreak, and before restocking.

An effective biosecurity plan approved by PDA will be required for issuance of a PDA permit for a flock of any size to move product if a flock is located in a disease control area. Since 2022, thousands of biosecurity plans have been reviewed in Pennsylvania for the issuance of permits to allow movement of birds, eggs, and manure. Written biosecurity plans must be submitted for approval every two years.

All NPIP participants in PA, regardless of flock size, must address biosecurity to have a successful on-site NPIP inspection.

All flock premises should have a biosecurity plan in place, and all people entering the premises should be trained in the plan requirements, and should follow that plan. The National Poultry Improvement Plan (Program Standard E) outlines the major elements for an acceptable biosecurity plan, and the PDA Biosecurity Evaluation Form provides details for each of the 14 items required. The plan template and evaluation form are available upon request to assist with development of a biosecurity plan. Biosecurity plan specialists are also available to assist with plan development and evaluation.

Biosecurity plans submitted for review are required to be written in the format specified by the NPIP Program Standard E template.

NPIP Program Standards Biosecurity Principles

Biosecurity Plan Template

2025

1. Biosecurity responsibility

1.1. Is there a Biosecurity Coordinator? If so, please provide their name.

1.2. Is there a site-specific biosecurity plan?

1.2.1. Although a company plan may be used for all premises under that company, the plan must be individualized for each premises, with a premises map included. The plan should also include the species, production type, and flock size of the premises.

1.3. Is the Biosecurity Coordinator knowledgeable in the principles of biosecurity?

1.3.1. The Biosecurity Coordinator must be able to describe and interpret their company's biosecurity program and how it meets the requirements of the NPIP biosecurity principles. PDA expects that the biosecurity coordinator listed on the plan is able to meet these requirements.

1.4. Does the Biosecurity Coordinator review the biosecurity plan at least once during each calendar year and make revisions as necessary?

1.4.1. The Biosecurity Coordinator must be able to provide recorded dates in which annual reviews were made and evidence of revisions to the biosecurity plan if any were necessary. These dates should be written in the biosecurity plan or on supporting documents which are provided along with the plan. Review of the plan by officials is only required every 2 years but the documentation should support that it was reviewed annually and/or when risk is elevated (see 1.5).

1.5. Does the biosecurity plan indicate there will be a review by the Biosecurity Coordinator in periods of heightened risk of disease transmission?

1.5.1. Documentation of compliance, including evidence of a discussion during periods of heightened risk can take any form (e.g., emails, letters, memos, phone logs, text messages, etc.). It is the responsibility of the Biosecurity Coordinator to clearly define and communicate the "period of heightened risk" in the biosecurity plan.) For PDA review, if there are items in the plan that are put into play only during times of higher risk, the plan should mention what these items are and what would be the trigger for them to be implemented. The plan should include how and when this is communicated to employees.

2. Training

2.1. Does the biosecurity program include training materials that cover both farm site-specific procedures as well as company and/or complex-wide site-specific procedures, as applicable?

All standard operating procedures (SOPs) for biosecurity should be listed by title in the plan and should be available to PDA upon request. Review of the SOPs should be included in initial and annual employee training. These SOPs could include the company/site SOPs for PPE, pest control, equipment sharing, C&D, etc. The training should include review of all SOPs and a review of the biosecurity plan and the plan should include this.

2.2. Do all poultry owners and caretakers that regularly enter the perimeter buffer area (PBA) complete this training?

2.2.1. Supporting documentation (e.g., training logs, training completion sheets, sign-in sheets, certificates of completion, etc.) should be provided.)

The plan should include that everyone who regularly crosses the PBA receives biosecurity training (as per 2.1).

2.3. Has the training been completed at least once per calendar year and documented?

2.3.1. Supporting documentation (e.g., training logs, training completion sheets, sign-in sheets, certificates of completion, etc.) should be provided.)

PDA requires that training logs are provided along with the plan at the initial plan evaluation and at evaluation every 2 years, to provide proof that all employees (and owners) initial and annual biosecurity training for the premises. This should include the name of the trainee and the date the plan was reviewed with them.

2.4. Are new poultry caretakers trained at hire?

2.4.1. Supporting documentation (e.g., training logs, training completion sheets, sign-in sheets, certificates of completion, etc.) should be provided.)

PDA requires that training logs are provided along with the plan at the initial plan evaluation and at evaluation every 2 years, to provide proof that all employees (and owners) initial and annual biosecurity training for the premises. This should include the name of the trainee and the date the plan was reviewed with them.

2.5. Are training records retained as stated in Title 9-CFR §145.12(b) and 146.11(e)?

2.5.1. Records must be maintained for at least 3 years.)

The plan should state that the training logs are kept for a minimum of 3 years.

3. Line of Separation (LOS)

3.1. Does the site-specific biosecurity plan describe or illustrate the boundaries of the LOS? If not, please explain.

3.1.1. Provide a diagram, map, and/or a detailed description of the LOS. The LOS is recommended but not required for poultry with unenclosed outdoor access but is highly recommended during periods of heightened risk.

The LOS should be drawn and labelled on a premises map (or described), and requirements for crossing the LOS should be included in the plan for each premises. For outdoor birds, the LOS should be the fence line and the entry point should be the gate.

3.2. Does the site-specific biosecurity plan clearly outline procedures to be followed when caretakers, visitors, or suppliers cross the LOS?

3.2.1. Provide the procedures (e.g., written instructions, signage, training videos, etc.) to be followed by caretakers, visitors or suppliers to cross the LOS.) The requirements for crossing the LOS should be included in the plan for each premises. For the LOS, biosecurity signs at the LOS entry at each house are strongly recommended but not required if there is signage at the PBA entry. The LOS procedures should include an entry point at the LOS clearly indicated by a bench, taped or painted line or other method of separation, and designated footwear or footwear which has been cleaned and disinfected before crossing the LOS. A foot pan is not acceptable as the only requirement to cross the LOS- but may be used in addition to the designated or footwear which was cleaned and disinfected. If foot pans are used, the plan should address that they are kept clean and disinfectant is replaced at least as often as the manufacturer recommends. Hand sanitizer or a hand washing station should be available and should be required for all. All employees and visitors should be aware of the requirements for crossing the LOS by signage or other methods.

4. Perimeter Buffer Area (PBA)

4.1. Does the site-specific biosecurity plan describe or illustrate the boundaries of the PBA?

4.1.1. Provide a diagram, map, and/or a detailed description of the PBA.

The PBA should be drawn and labelled on a premises map (or described), and requirements for crossing the PBA should be included in the plan for each premises. Structures and composting sites/disposal sites; feed bins; traffic flow; C&D station; entrance/exit should be labelled.

4.2. Does the site-specific biosecurity plan clearly outline the procedures to be followed by caretakers, visitors, or suppliers when entering and leaving the PBA?

4.2.1. Provide the procedures (e.g., written instructions, signage, training videos, etc.) to be followed by caretakers, visitors or suppliers when entering and leaving the PBA.)

The requirements for crossing the PBA should be included in the plan for each premises. The plan should include biosecurity/no trespassing signs at the entrance to the PBA, along with a C&D station (if necessary for vehicles without on-board disinfection sprayers), parking requirements/locations, and any requirements for PPE as they cross the PBA for employees, visitors, and for truck drivers.

5. Personnel

5.1. Does the biosecurity program and/or site-specific biosecurity plan include provisions specifically addressing procedures and biosecurity personal protective equipment (PPE) for site-dedicated personnel?

5.1.1. PPE should be described in the biosecurity plan for each type of production facility.

This section should include all the PPE or clean/dedicated clothing and footwear requirements for all employees on the site, including the requirements for crossing the PBA and the LOS. The plan should include any protocols for cell phones and other items taken across the LOS.

5.2. Does the biosecurity program and/or site-specific biosecurity plan address the procedures and biosecurity PPE for non-farm personnel?

5.2.1. PPE should be described in the biosecurity plan for each type of production facility for non-farm personnel.

This section should include all the PPE or clean/dedicated clothing and footwear requirements for all visitors on the site, including those for crossing the PBA and the LOS. Biosecurity protocols for vendors such as catch crews and vaccination crews should be on file with PDA if the plan under review refers to those vendor plans. PPE should be made available for any visitor not included under a vendor's biosecurity plan/PPE. Truck drivers should not exit their trucks without clean or disposable boots and should avoid crossing the LOS unless necessary for delivering or catching birds.

5.3. Does the biosecurity program and/or site-specific biosecurity plan specify procedures which all personnel having had recent contact with other poultry or avian species should follow before re-entering the PBA?

5.3.1. Supporting documentation (e.g., signed statements, acknowledgement forms, visitor log-in, signed policy documents, etc.) should be provided.)

Personnel and visitors should be prohibited from contact with other avian species for a minimum of 48 hours. The visitor log should include a statement for visitors to document that they have not been around avian species for at least 48 hours. It should also include the following:

- | |
|--|
| <ul style="list-style-type: none">a) visitor's nameb) company or affiliationc) reason for visitd) datee) if the visitor entered the LOSf) reason for entering LOS |
|--|

6. Wild Birds, Rodents and Insects

6.1. Are there control measures in the biosecurity program and site-specific biosecurity plan to prevent contact with and protect poultry from wild birds, their feces and their feathers as appropriate to the production system?

The plan should include steps taken to prevent wild bird access to poultry areas, including inlet wire, fan wire or shutters, etc. For outdoor birds, the plan can include any netting if used, fencing, and any measures taken to preclude entry of wild birds, such as clean up of any spilled feed, wild bird deterrents used on ponds, etc.

6.2. Does the biosecurity program and site-specific biosecurity plan contain control programs for rodents, insects, and other animals?

The plan should include any control programs for rodents, insects (usually flies), and should include that pets and other animals are prevented access to the poultry areas (should not cross LOS).

6.3. Are these programs documented?

6.3.1. Provide description of control programs and examples of the documentation [e.g., log sheets, rodent control company contracts, Best Management Practices (BMP) audits, maintenance records, etc.].

The plan should indicate that rodent and insect control programs are documented (counts, methods, bait rotations, etc). Documentation of control programs should be available upon request, but these logs are not required to be submitted with the plan.

7. Equipment and Vehicles

7.1. Does the biosecurity program and/or site-specific biosecurity plan include provisions for procedures for cleaning, disinfection, or restriction of sharing of equipment where applicable?

7.1.1. Supporting documentation (e.g., written instructions, signage, training videos, etc.) should be provided.)

The plan should address the policies for equipment sharing, and if equipment is shared, address the procedures used for drivers, and C&D of any equipment shared. This section should mention procedures used

for all vehicles and equipment crossing the PBA if not addressed in the PBA section. Supporting documentation is not required at the time of plan evaluation; however, should be available upon request. Any SOPs should be listed by title in the plan.

7.2. Are vehicle access and traffic patterns defined?

7.2.1. Provide a description of vehicle entry access and traffic patterns.) Include the location of the C&D station on the site map.

The site map should include the location of the C&D station, or if all vehicles entering the PBA use on-board sprayers, the location where these sprayers are activated. If this is not included on the map, it should be described in the written plan. The map should include arrows showing traffic patterns unless described in the written plan.

8. Mortality Disposal

8.1. Is there a mortality disposal plan?

This addresses a plan for routine daily mortalities.

8.2. Does the mortality disposal plan reference the frequency of removal, storage of mortality, and pest control around mortality storage and disposal areas?

8.2.1. Provide a description of the mortality disposal plan and examples of documentation [e.g., mortality sheets, company contracts, Best Management Practices (BMP) audits, disposal records, etc.].

The plan should include a description of daily mortality management, including frequency of removal from the poultry house or area; method of disposal (ie-composting on site); location of disposal site; method of moving mortality from the poultry area to the disposal site in a biosecure manner (ie- is any equipment crossing the LOS or PBA, and are any personnel crossing the LOS or PBA to dispose of mortality). The protocol for C&D of any equipment, including buckets used for moving mortalities, should be included in the plan. Supporting documentation is not required at the time of plan evaluation; however, should be available upon request. Any SOPs should be listed by title in the plan.

8.3. Does the mortality disposal plan address procedures for handling mortality disposal in a way that minimizes the potential for cross-contamination from other facilities or between premises?

8.3.1. Supporting documentation should be provided (e.g., written instructions, videos, etc.) for proper handling of mortality to minimize the potential of cross-contamination.)

The plan should address the management of daily mortalities, equipment, transport, and personnel if the mortalities are transported off of the premises. (ie- C&D of vehicle and other equipment used, driver PPE, etc). The plan can refer back to other sections addressing the equipment and personnel. Supporting documentation is not required at the time of plan evaluation; however, should be available upon request. Any SOPs should be listed by title in the plan.

If using landfill for disposal, carcasses of birds which have died within 48 hours of pickup may be bagged in heavy duty garbage bags and kept in a dumpster for pickup. Carcasses of birds which died 48 hrs or longer before pickup must be kept in a freezer before pickup.

Dumpsters should be kept clean, especially of leakage of fluids from carcasses, and maintained in an area away from the poultry area. Rodent control should be in place around the dumpster.

Barrels and buckets used to hold mortality temporarily should be tightly covered.

9. Manure and Litter Management

9.1. Is the manure and spent litter handled in a manner that limits the spread of infectious disease?

9.1.1. Procedures (e.g., written instructions, manure/litter handling log sheets, protocols, permits, guidance for contractors, etc.) should be provided showing how disease risk from manure and litter handling has been addressed.)

The plan should address the management of manure/spent litter, equipment, transport, and personnel if it is transported off of the premises. (ie- C&D of vehicle and other equipment used, driver PPE, etc). The plan can refer back to other sections addressing the equipment and personnel. Supporting documentation is not required at the time of plan evaluation; however, should be available upon request. Any SOPs should be listed by title in the plan.

10. Replacement Poultry

10.1. Is replacement poultry sourced from flocks which are in compliance with NPIP provisions and program standards?

10.1.1. Replacement poultry is defined as poultry from hatch to maturity intended to become laying hens or breeders, but per USDA in 2022, all poultry on the premises should be sourced from NPIP sources or sources which tested equivalently to NPIP standards to be eligible for indemnity (or the affected flock is tested equivalently). However, PDA may allow permitting relative to a disease control area if this requirement is not met.)

(10.1.2. Provide supporting documentation (e.g., VS 9-2 form, VS 9-3 form and/or NPIP hatchery production records) showing that source flocks are active and compliant participants in the NPIP.)

9-3 forms should be obtained from NPIP bird sources and should be available at the time of plan evaluation, or a 9-2 form may be completed for NPIP participants when birds are placed may be used instead. Alternatively, an invoice showing the flock source and NPIP number may be substituted. ALL FLOCKS BEING EVALUATED FOR INDEMNITY PURPOSES MUST HAVE PROOF THAT THEIR FLOCKS ORIGINATED FROM AN NPIP BREEDER FLOCK OR FROM A FLOCK THAT WAS TESTED EQUIVALENTLY TO NPIP (Per NPIP Director 11/16/22).

10.2. Is replacement poultry transported in equipment and vehicles that are regularly cleaned, disinfected and inspected?

10.2.1. Supporting documentation (e.g., written instructions, wash station reports and/or logs, inspection reports, invoices, etc.) should be provided. The biosecurity plan should be followed as written and clearly define “regular cleaning.”

In many cases, unless using their own company trucks, this will be covered by the poultry transport companies’ biosecurity plans. These plans should be on file with PDA at the time of plan evaluation. Supporting documentation is not required at the time of plan evaluation; however, should be available upon request. Any SOPs should be listed by title in the plan.

10.3. Are biosecurity protocols in place for equipment and personnel involved in the transport of replacement poultry?

10.3.1. Supporting documentation [e.g., signed statements, acknowledgement forms, visitor log-in sheets, policy documents, standard operating procedures (SOPs), Best Management Practices (BMPs), etc.] should be provided for personnel involved in the transport of replacement poultry.)

10.3.2. Supporting documentation [e.g., written instructions, protocols, procedures, training videos, standard operating procedures (SOPs), Best Management Practices (BMPs), etc.] should be provided for equipment involved in the transport of replacement poultry.

In many cases, this will be deferred to the poultry transport companies’ biosecurity plans, which should be on file with PDA at the time of plan evaluation. All personnel, vehicles and equipment entering the premises must meet the premise’s biosecurity requirements before crossing the PBA and all personnel and equipment must meet the premise’s biosecurity requirements before crossing the LOS. Supporting documentation is not required at the time of plan evaluation; however, should be available upon request. Any SOPs should be listed by title in the plan.

11. Water Supply

11.1. Is drinking water and/or water used for evaporative cooling sourced from a contained supply such as a well or municipal system?

11.2. If water comes from a surface water source, is water treatment used to reduce the level of disease agents?

11.2.1 If yes, describe the water treatment used.

11.3 If water treatment is not possible, is a risk analysis performed to determine actions needed to mitigate risks?

11.3.1 The Biosecurity Coordinator should provide evidence that the risk of an untreated system has been considered and demonstrate steps to mitigate that risk if feasible.

11.3.2 Risk assessment should have considered risks from the water supply. This guidance does not require a peer-reviewed professional risk assessment to be performed.

11.4 If surfaces have been cleaned or flushed with surface water, is subsequent disinfection employed to prevent disease transmission?

11.4.1 If surfaces were cleaned or flushed with surface water and subsequent disinfection was employed, a description of the subsequent disinfection and/or supporting documentation should be provided (e.g., invoices for chemicals used and purchased treatment equipment, treatment plans, etc.)

11.4.2 If surfaces were cleaned or flushed with surface water and subsequent disinfection was not employed, see item 11.3.2 above.

12. Feed and Replacement Litter

12.1. Are feed, feed ingredients and litter stored and maintained in a manner that limits exposure to and contamination by wild birds, rodents, insects, and other animals?

12.1.1. Guidance applies to the feed ingredients and litter which are under the direct control of the entity being audited. A description or examples (e.g., written instructions, feed or replacement litter handling, log sheets, protocols, permits, guidance for contractors, etc.) should be provided showing how exposure to and contamination by wild birds, rodents, insects, and other animals is limited. Occasional exceptions may be necessary (e.g., seasonal storage, “acts of God”, etc.).

12.1.2. Guidance applies to the feed ingredients and litter which are under the direct control of the entity being audited.

This premise’s biosecurity plan should address the storage and handling of feed and fresh litter after it has been delivered.

12.2. Does the biosecurity plan address feed spills within the PBA (outside of the LOS)?

The plan should include frequency of clean up for feed spills.

13. Reporting of Elevated Morbidity and Mortality

13.1. Does the biosecurity plan address elevated morbidity and/or mortality above expected levels?

13.1.1 A description of the actions and/or documentation (e.g., evidence of investigation, tracking graphs, mortality/morbidity patterns, case reports, mortality logs, etc.) used to monitor morbidity and/or mortality should be provided. The Biosecurity Coordinator is responsible for communicating what constitutes elevated morbidity and/or mortality in the biosecurity plan.

The plan should address the reporting of elevated morbidity and mortality, and what parameters are used to determine if these are elevated.

13.2. Is there a plan to report to responsible authorities and take appropriate action should you suspect and need to rule out reportable disease agents?

13.2.1. Provide the written procedure to report and take appropriate actions when disease agents are suspected. The Biosecurity Coordinator is responsible for providing the written procedures.

13.2.2. The written procedure should identify the responsible reporting authorities. The Biosecurity Coordinator is responsible for clearly communicating who the responsible authorities are.

The plan should include mention of reporting suspicions of HPAI to PDA and should include the PDA 24/7 number. 717-772-2852 (option 1)

14. Auditing

14.1. Auditing of the biosecurity principles is based on flock size as outlined in 9 CFR 53.10.

14.1.1. Premises exempted from auditing for USDA indemnity purposes are those which: raise fewer than 100,000 broilers annually for meat, raise fewer than 30,000 turkeys annually for meat, have fewer than 75,000 table egg layers, raise fewer than 75,000 pullets annually; raise fewer than 25,000 raised for release upland game birds or waterfowl annually; raise fewer than 25,000 egg-type upland game birds and waterfowl annually, or have fewer than 5,000 breeders. All flocks require a biosecurity plan for a permit to restock birds in a control area to be approved by PDA.

14.2. Audits shall be conducted at least once every two years or a sufficient number of times during that period by the Official State Agency to ensure the participant is in compliance.

14.2.1. Audits conducted by PDA shall be a paper-based assessment of the participant's biosecurity plan. On farm assessments are offered and required for commercial flocks restocking in a control area.

PDA inspectors will complete a biosecurity risk assessment for any poultry producer who wants to assess and improve their biosecurity plans on site. Call PDA or your regional PDA veterinarian to discuss an assessment for your farm.

Biosecurity Plan Line of Separation and Perimeter Buffer Area Example

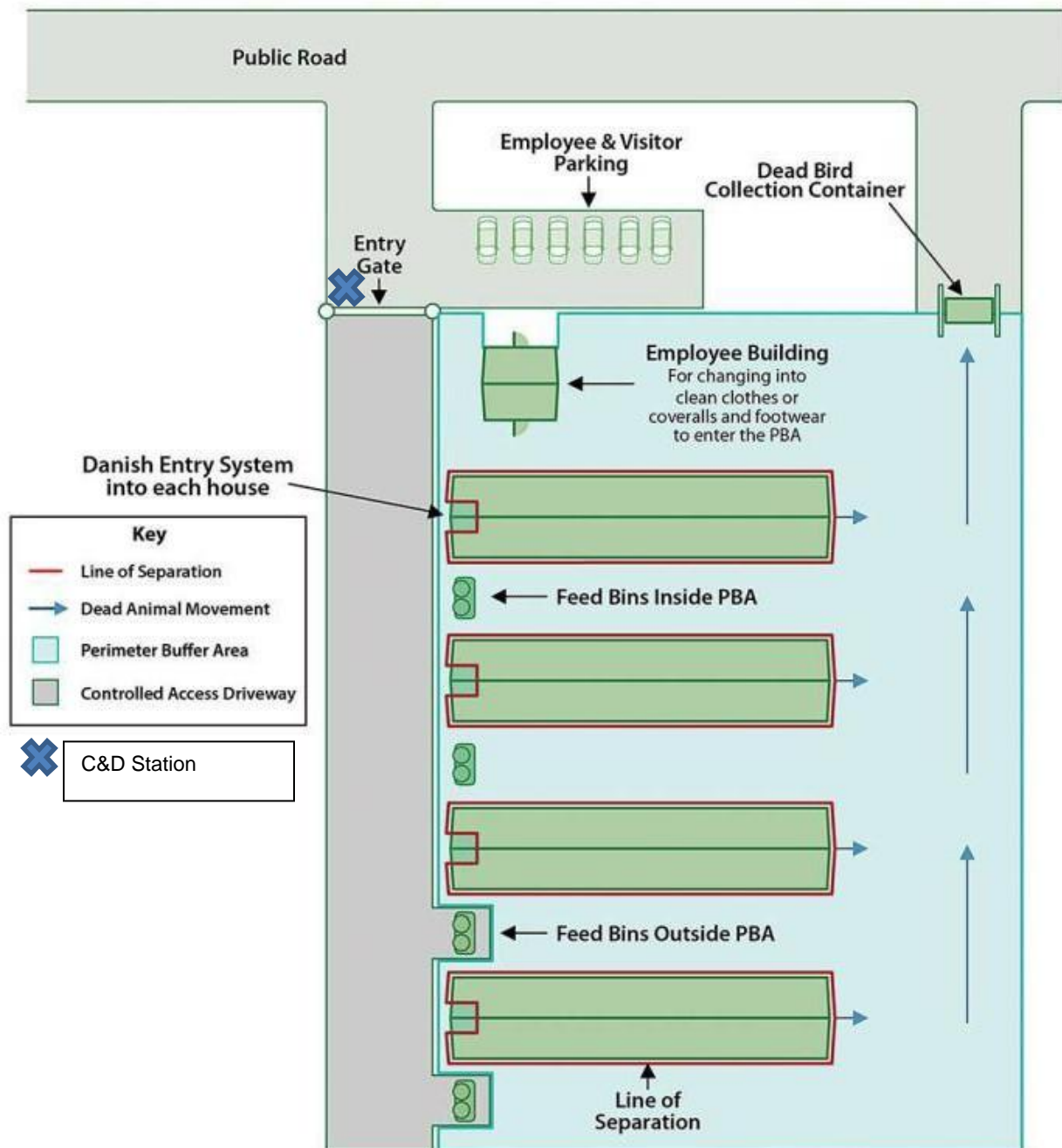


Figure courtesy of the Center for Food Security and Public Health, Iowa State University

SECTION VIII: PREMISES REGISTRATION FORM

PA PREMISES DATA INFORMATION SHEET

Address:

County: Township:

Primary contact (Correspondence and Emergency Contact)

Name: Address:

Home Phone: Business Phone: Cell Phone:

Check all that apply: ☐ Land Owner ☐ Manager ☐ Animal Owner ☐ Other:

Email:

Secondary contact (correspondence and Emergency Contact)

Name: Address:

Home Phone: Business Phone: Cell Phone:

Check all that apply: ☐ Land Owner ☐ Manager ☐ Animal Owner ☐ Other:

Domestic Species at this location?

(Check all that apply & list number)

<input type="checkbox"/> Dairy Cattle <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Heifer Grower <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Veal Grower <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Beef Cattle <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Cow/Calf <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Backgrounder <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Feedlot <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Swine <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Grower <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Nursery <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Finisher <input style="width: 60px; height: 20px;" type="text"/>	<input type="checkbox"/> Sheep * <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Goats * <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Dairy <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Meat <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Camelids <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Alpaca <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Llama <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Cervids <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Deer <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Elk <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Equine <input style="width: 60px; height: 20px;" type="text"/>	<input type="checkbox"/> Turkeys <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Waterfowl <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Ratite <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Upland Game Birds <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Pigeons <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Chickens <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Egg <input style="width: 60px; height: 20px;" type="text"/> <input type="checkbox"/> Meat <input style="width: 60px; height: 20px;" type="text"/>
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***If you have a USDA Scrapie ID please list**

Return Form to: Bureau of Animal Health & Diagnostic Services
2301 N. Cameron St. | Harrisburg, PA 17110-9408 | Ste. 412 | 717.772.2852 | www.agriculture.pa.gov

SECTION IX: PDA CPT QUARANTINE ORDER

NOTICES

DEPARTMENT OF AGRICULTURE

General Quarantine Order; Certified Poultry Technicians Revised 2025

[55 Pa.B. 2425]

[Saturday, March 29, 2025]

Recitals.

A. The Act of April 6, 1956, P.L. (1955) 1429, as amended February 6, 2025, P.L. 1, No. 1 (CPT Act), provides, at section 641, that "The Secretary of Agriculture, when he determines that there is a need for trained technicians to aid in the collection of regulatory samples from poultry and eggs and conduct on-site surveillance for dangerous transmissible diseases of poultry, may license as many certified poultry technicians as he deems necessary to carry out specified regulatory sampling and testing procedures. The Secretary of Agriculture may establish the licensing and certification standards, and the regulatory sampling and testing procedures, and may provide training in Spanish, through the issuance of a general quarantine order under the authority of 3 Pa.C.S. § 2329 (relating to quarantine)." (63 P.S. § 641)

B. The Pennsylvania Department of Agriculture (Department), under authority of section 2329 of the Domestic Animal Law (3 Pa.C.S. § 2329) (related to quarantine) has the power to establish and enforce a quarantine order "Whenever a dangerous transmissible disease. . . exists anywhere within or outside of this Commonwealth, or whenever it is deemed advisable to test or treat any domestic animal upon the reasonable suspicion that it has contracted or been exposed to a dangerous transmissible disease. . . or whenever the testing or treatment of a domestic animal indicates that the domestic animal has been exposed to a dangerous transmissible disease. . . so as to render future accurate testing for recent exposure of that domestic animal to that dangerous transmissible disease. . . impractical or impossible, the department shall have the power to establish and enforce quarantines of any such infected, exposed, contaminated, suspected or susceptible domestic animal."

C. The Secretary of Agriculture has determined that there is a need for trained technicians to aid in the collection of regulatory samples from poultry and eggs and conduct on-site surveillance for dangerous transmissible diseases of poultry.

D. Through the following Order, the Secretary establishes the licensing, training and certification standards and the regulatory sampling and testing procedures required for certification and licensing of Certified Poultry Technicians.

Order.

PDA enters a General Quarantine Order, incorporating the foregoing recitals. This Order is entered under authority of the Domestic Animal Law (at 3 Pa.C.S. § 2329), the CPT Act (63 P.S. §§ 641-647) and § 1702 of the Administrative Code of 1929 (71 P.S. § 442), and establishes the standards with respect to training and testing of persons to become licensed as Certified Poultry Technicians and provides the sampling and testing procedures and standards for the collection of regulatory samples for testing and rapid testing for regulatory purposes for dangerous transmissible diseases of poultry.

1. *General.* For purposes of this General Quarantine Order, poultry shall include, chickens, domestic waterfowl, turkeys, pigeons, doves, pheasants, grouse, partridges, quail, guinea fowl, pea fowl, gallinaceous birds, and ratites, of any age, that may contract or harbor Avian Influenza virus whether living or dead.

2. *Purpose.* This Order allows for qualified persons to become licensed as Certified Poultry Technicians to carry out certain regulatory sampling and testing procedures, and thereby further fulfill the Department's duty to surveil for dangerous transmissible diseases in accordance with the Domestic Animal Law. This Order allows for a more proactive and preventative approach to current and future dangerous transmissible diseases and hazardous substances through surveillance, monitoring, and testing by the poultry industry. Such an approach furthers the legislative mandate of section 2327 of the Domestic Animal Law (3 Pa.C.S. § 2327) (related to disease surveillance and detection) and provides greater protection to the poultry population and industry in this Commonwealth. This Order delineates education, regulatory sampling, and testing criteria that will allow for and require the certification and licensure of private sector individuals to carry out domestic poultry sampling and rapid testing and thereby enhance monitoring, detection, and containment of dangerous transmissible diseases of poultry and contamination by hazardous substances.

3. *Scope of certification and licensure.* This Order allows persons licensed as Certified Poultry Technicians in the Commonwealth of Pennsylvania to perform the duties of a Certified Poultry Technician only within the Commonwealth of Pennsylvania, unless under a reciprocal agreement addressing license reciprocity between the Commonwealth of Pennsylvania and another state or Commonwealth. Final certification shall be obtained, and the certificate issued prior to any person undertaking any testing, sampling or other activity requiring certification and licensure under this Order.

4. *Qualifications for certification and licensure.*

(a) Nothing in this Order may or shall supplant the provisions of the Veterinary Medicine Practice Act (Act of December 27, 1974, P.L. 995, No. 326, as amended) (63 P.S. § 485.1 et seq.) or its attendant regulations regarding the practice of veterinarian medicine. Persons licensed as Certified Poultry Technicians may only carry out those testing and diagnostic practices that would not rise to the level of veterinary medical practice as defined under the Veterinary Medicine Practice Act.

(b) Applicants for licensure as a Certified Poultry Technician shall furnish such information as the Department may reasonably require in order to ascertain competence and qualification, including at a minimum the following information and evidence thereof:

- (i) That the applicant is at least eighteen (18) years of age;
- (ii) That the applicant can legally work in the United States in accordance with Federal and State law;
- (iii) That the applicant has satisfactorily completed a course of training as a Certified Poultry Technician as prescribed by this Order; and

(iv) An attestation related to whether the applicant has ever been convicted of a violation of the Domestic Animal Law, a felony, or a cruelty to animals charge under the Pennsylvania Crimes Code within the time periods established in subsection (c) below.

(c) An application for certification and licensure as a Certified Poultry Technician, including an application for renewal of license, shall be denied if the applicant has done any of the following:

(i) Has been convicted, within the previous three years of any provision of the Domestic Animal Law (3 Pa.C.S. § 2301 et seq.), the CPT Act (63 P.S. §§ 641-647), or any regulation promulgated or order issued thereunder by the Department.

(ii) Within the previous ten years the applicant has been convicted of a felony, including a felony conviction for an offense under 18 Pa.C.S. Chapter 55 Subchapter B (relating to cruelty to animals).

(iii) Within the previous three years the applicant has been convicted of an offense under 18 Pa.C.S. Chapter 55 Subchapter B (relating to cruelty to animals) or of substantially similar conduct pursuant to a cruelty law in another state or Commonwealth.

(d) Persons licensed by the Department as Certified Poultry Technicians under authority of the “General Quarantine Order; Certified Poultry Technicians” [49 Pa.B. 3088, Saturday, June 15, 2019] , may continue to carry out the duties and functions bestowed upon them under that Quarantine Order only until such time as that license has expired or been revoked in accordance with the provisions of that Quarantine Order.

(e) Upon expiration of a Certified Poultry Technician license issued under the “General Quarantine Order; Certified Poultry Technicians” the person shall obtain the certification and licensure required by this Order. Failure to do so shall mean the person may no longer perform the duties and functions of a Certified Poultry Technician.

5. *Application for certification and licensure.*

(a) A completed Certified Poultry Technician License Application shall be submitted to the Department for review before acceptance into the certification and training course.

(b) An applicant for Certified Poultry Technician certification and licensure shall complete the required and appropriate certification course and training, including passing a written or examination, and subsequent field skills testing, administered or approved by the Department. Field skills testing must be successfully completed within six months after the classroom course and examination has been successfully completed.

(c) After successful completion of the field skills testing, submission of the form evidencing successful completion of the field skills testing to the Department shall be required within thirty (30) days after the date of the testing. Upon receipt of the completed field skills testing form, the Department will mail or email the license to the approved applicant. The applicant may then begin performing the duties of a Certified Poultry Technician.

6. *Annual certification license; Renewal application for licensure.*

(a) Annual license. A license for a Certified Poultry Technician shall expire on January 1 of each calendar year.

(b) Renewal. A license shall be renewed on an annual basis. Applications for renewal shall be submitted to the Department and postmarked prior to January 1 of the year for which renewal is sought.

(c) Failure of a licensed Certified Poultry Technician to submit a license renewal application postmarked prior to January 1 of the year in which renewal is due will result in the following:

(i) The person will be unlicensed to practice as a Certified Poultry Technician.

(ii) The person may not continue to act or hold themselves out as a licensed Certified Poultry Technician.

(iii) Continuing to act or hold oneself out as a licensed Certified Poultry Technician shall be a violation of this Order and shall subject the person to the penalties provided for at sections 2383 (related to enforcement and penalties) and 2386 (related to civil remedy) of the Domestic Animal Law (3 Pa.C.S. §§ 2383 and 2386), and to penalties set forth under section 647 of the CPT Act (63 P.S. § 647).

(iv) The person shall be required to successfully complete the written examination at a Department office, and, at the Department's discretion, may be required to complete the entire certification course, examinations and training as established in Sections nine (9) and 10 of this Order prior to the Department issuing a license.

7. Duties and Limitations.

(a) Any person licensed as a Certified Poultry technician may engage in carrying out the duties and procedures within the scope of their certification and licensure as outlined under the provisions of this Order.

(b) A licensed Certified Poultry Technician may carry out the following:

(i) The collection of blood, swab, and egg samples of sufficient quantity and quality for regulatory testing from poultry in accordance with applicable statutory and regulatory standards, and orders of the Department;

(ii) Performance of rapid tests for regulatory testing as approved by the Department and in accordance with applicable statutory and regulatory standards, and orders of the Department;

(iii) Handling, packaging, labeling, and timely submission of samples in accordance with applicable statutory and regulatory standards, and orders of the Department;

(iv) Completion of sample submission forms and all other forms required by the Department or USDA;

(v) Application of official animal identification as approved by the Department.

(vi) Maintenance of records of animal identification, approved rapid testing, and any other records required by the Domestic Animal Law, this Order, and any order issued by the Department;

(vii) Timely reporting of any suspected reportable diseases in accordance with applicable statutory and regulatory standards, and orders of the Department; and

(viii) Sampling and testing to meet Commonwealth of Pennsylvania and federal avian program standards and Commonwealth exhibition standards.

(c) *No Veterinary Functions.* Being licensed as a Certified Poultry Technician shall not entitle the person to perform any function for which a veterinary license or certification as a veterinary technician is required.

(d) *Ongoing oversight.* The Department may observe any Certified Poultry Technician performing the duties of a Certified Poultry Technician to assure compliance with the provisions of the Domestic Animal Law and this Order, and to assure proper procedures and protocols are being instituted and followed and may conduct sampling and testing of its own to verify and assure the accuracy of the sampling and testing being conducted by a Certified Poultry Technician.

8. Certification Requirements.

(a) *General.* The Department will develop the Poultry Technician certification course in accordance with the standards established in Sections nine (9) and 10 of this Order. The Department may administer the course or may approve certification courses offered by an approved vendor. All certification courses shall be conducted by either a Department veterinarian or a designee of the Department with the oversight of a Department veterinarian.

(b) *Access to Training.* The classroom portion of the certification course and examination will be offered at least once per calendar year, or more often as deemed necessary by the Department and will be offered at various locations across the Commonwealth in a manner that will assure reasonable Commonwealth-wide coverage and access.

(c) *Criteria.* The following establish the criteria related to certification and maintaining certification under this Order:

(i) A person seeking to act as a Certified Poultry Technician shall complete the necessary certification course work, field skill training, and evaluation, which shall include classroom instruction and testing and field skill instruction and testing. A person shall be required to obtain a minimum score of eighty percent (80%) on a classroom written examination and have all applicable skills checked as satisfactory on field skills testing.

(ii) A person acting as a Certified Poultry Technician shall maintain certification and licensure in accordance with the requirements set forth in this Order.

(iii) A person certified and acting as a Certified Poultry Technician shall conduct such sampling, testing, and other related activities in accordance with the requirements of this Order and any order issued by the Department.

(iv) A person certified as a Certified Poultry Technician shall maintain records of official animal identification, approved rapid testing, and any other records required by the Domestic Animal Law, this Order, and any order issued by the Department for a minimum of two years and shall make records available upon Department request.

9. Certification course and training.

(a) At a minimum, the successful completion of the certification course, training and examinations will demonstrate an applicant's understanding of and technical knowledge and proficiency relating to the duties as described in this Order. The examinations will address key topics and skills addressed during the classroom portion of the certification course and field skills training, as set forth in this Order.

(b) Field skills training will be offered to individual applicants at regional locations chosen by the Department to provide reasonable access and will be administered by a Department veterinarian or designee with the oversight of a Department veterinarian.

(c) The certification course, training and testing requirements for a Certified Poultry Technician shall, at a minimum, address proper methods for the following:

(i) The collection of blood, swab, and egg samples of sufficient quantity and quality for regulatory testing from poultry in accordance with applicable Pennsylvania and federal statutory and regulatory standards, Quarantine Orders of the Department and federal National Poultry Improvement Plan standards.

(ii) Performance of rapid tests for regulatory testing as approved by the Department and in accordance with applicable Pennsylvania and federal statutory and regulatory standards, Quarantine Orders of the Department and federal National Poultry Improvement Program standards.

(iii) Handling, packaging, labeling, and timely submission of samples and sample submission forms in accordance with Pennsylvania statutory and regulatory standards and Quarantine Order provisions.

(iv) Comprehensive, timely and accurate completion of sample submission forms and all other forms required by the Department or USDA.

(v) Requirements for official animal identification and application of such.

(vi) Maintenance of records of official animal identification, approved rapid testing, and any other records required by the Domestic Animal Law, its attendant regulations, this Order, and any Quarantine Order issued by the Department.

(vii) Timely reporting of any suspected reportable diseases in accordance with applicable Domestic Animal Law statutory and regulatory standards, Quarantine Orders of the Department and statutory, regulatory and program standards, such as National Poultry Improvement Program standards established by the USDA.

(viii) Knowledge of and adherence to requirements for sampling and testing to meet Pennsylvania and federal statutory and regulatory standards, Quarantine Orders of the Department and federal National Poultry Improvement Program standards.

(ix) Understanding of and adherence to Pennsylvania and federal statutory, and regulatory standards and federal program standards and indemnity requirements related to biosecurity measures and practices.

10. *Certification Examination.*

(a) *Results.* The Department will notify the applicant of the results of the written examination and field skills test.

(b) *Written examination.*

(i) If the applicant passes the written examination, the applicant may schedule field skills testing with a Department regional veterinarian or designee.

(ii) If the applicant fails the written examination the applicant shall be allowed to take the next available written examination offered within one year from the date of the failed examination without repeating the classroom portion of the training.

(iii) If the applicant fails the written examination for a second time the applicant shall be denied certification and the applicant shall be required to again complete all the necessary certification course training, pass the written examination and field skills testing, and obtain final certification under the provisions of this Order.

(iv) An applicant who has failed the written examination may request a copy of the results via by email or fax or by sending a written request, along with a self-addressed postage paid envelope, to the Department.

(c) *Field skills testing.*

(i) If the applicant passes the field skills testing, the Department instructor who evaluated the applicant will notify the applicant of the results.

(ii) In order to obtain certification, the applicant must remit a copy of the field skills testing form, signed by the instructor, to the Department's Harrisburg office. This submission shall be required within thirty (30) days after the date of the field skills testing.

(iii) If the applicant fails the field skills testing, the applicant shall be allowed to schedule up to two additional field skills tests within the six months after successfully completing the written examination without repeating the classroom training or written examination.

(iv) If an applicant fails the field skills testing for a third time, the applicant shall be denied certification and the applicant shall be required to again complete all the necessary certification course training, pass the written examination and field skills testing, and obtain final certification and licensure under the provisions of this Order.

11. *Recordkeeping.*

(a) *General requirements.* The Department will make available forms on which records of official animal identification, approved rapid testing, and any other records required by the Domestic Animal Law, this Order, and any order issued by the Department shall be recorded. Such forms shall be reviewed during the classroom training and shall be available from the Department for the regulated community.

(b) *Duty to keep and submit records.* A Certified Poultry Technician shall be responsible for keeping records as required by the Department, such as records of official animal identification, records of approved rapid testing performed and results of tests on each animal, and any other records required by the Domestic Animal Law, this Order, and any order issued by the Department. The Certified Poultry Technician shall keep required records for a minimum of two years and shall make records available upon Department request. Such records shall be kept on forms provided by the Department. All required records shall be made available to the Department for inspection or copying or both upon request of the Department.

12. *Duty to report.*

Consistent with the purpose of the Domestic Animal Law and the provisions established at section 2327 (related to disease surveillance and detection) (3 Pa.C.S. § 2327(b)), a Certified Poultry Technician shall, immediately upon receiving information thereof, report to the Department each case of any dangerous transmissible disease and each case of potential contamination by substances declared hazardous by the Department. Failure to report shall be considered a violation of this Order.

13. *Reciprocity.*

(a) *General.* A person who has a valid certificate or license from another state or Commonwealth may obtain licensure in this Commonwealth if:

(i) The state or Commonwealth in which that person is certified or licensed has a reciprocal agreement with the Department.

(ii) The training requirements in the state or Commonwealth from which the applicant is applying satisfy and are at least as stringent as the required certification and licensing training standards set forth in this Order.

(iii) The applicant is currently certified or licensed and is in good standing in the state or Commonwealth with which the Department has a reciprocal agreement.

(iv) The applicant meets the qualification standards established in this Order.

(b) *Procedure.* A person desiring a license under sections five (5) and six (6) (relating to application for certification and licensure) shall submit to the Department a properly completed application as set forth in this Order, along with a copy of the person's out-of-State certificate or license and an affidavit or verification of good standing signed by the appropriate regulatory body in that state or Commonwealth.

(c) *Jurisdiction.* An out-of-state person applying for and receiving certification and licensure to act as a Certified Poultry Technician in the Commonwealth of Pennsylvania shall be subject to the Jurisdiction of the Department and this Order and shall be subject to all penalties and requirements established in the Domestic Animal Law and this Order.

(d) *Other state.* A Certified Poultry Technician certified in the Commonwealth of Pennsylvania wishing to perform the duties of a Certified Poultry Technician in another state or Commonwealth may only do so after meeting the application and certification requirements of a state or Commonwealth with which a reciprocity agreement exists.

14. *Violations and Penalties.*

(a) *Denial, suspension and revocation of license.* The Department may, after notice, including a statement of the reasons therefore, deny, suspend or revoke the license of a Certified Poultry Technician for any of the following:

(i) A violation of the Domestic Animal Law, the CPT Act, this Order or any order, rule or regulation issued by the Department.

(ii) Failure of a Certified Poultry Technician to meet the requirements of certification and licensure.

(iii) Inconsistency and demonstration of a lack of knowledge in the skills and techniques necessary to carry out the duties of a Certified Poultry Technician.

(iv) Deficiencies in sample collection, sample handling and submission, and testing techniques, procedures, requirements and criteria established by the Department.

(v) Failure to renew certification and licensure in the established timeframe.

(vi) Being unwilling or unable to carry out the duties of a Certified Poultry Technician.

(vii) Falsifying information, including on applications, reports, records or correspondence with the Department.

(viii) A violation of any requirement of certification and licensure, recordkeeping or other provision of this Order.

(ix) Denial, suspension or revocation of certification or licensure in a state that has a reciprocal agreement with the Department.

(x) The Department may impose a civil fine or revoke, suspend or deny, or both, the license of a Certified Poultry Technician for any conviction of an offense under 18 Pa.C.S. Chapter 55 Subchapter B (relating to cruelty to animals), or substantially similar conduct pursuant to a cruelty law of another state or Commonwealth and shall revoke the license of a Certified Poultry Technician where such person has been convicted of a felony under any statute in this Commonwealth or another state or Commonwealth.

(xi) It shall be unlawful for any person to impede, hinder or interfere with the sampling or testing of a domestic animal or to refuse to confine a domestic animal so as to allow testing without undue burden on the official conducting the test or to fail to present the person's domestic animals for testing by the Department under authority of this Order after reasonable notice of the proposed testing has been given.

(xii) It shall be unlawful for any person who has knowledge that a domestic animal is infected with a dangerous transmissible disease or has been exposed to a dangerous transmissible disease or has been contaminated by a hazardous substance to conceal or attempt to conceal such a domestic animal or knowledge of such a domestic animal from the department.

(xiii) It shall be unlawful and a certification may be suspended, denied or revoked or civil fines and criminal penalties imposed, or any of the above, if a Certified Poultry Technician has violated any provision of the Domestic Animal Act or a final order of the Department, including failure to pay a civil penalty or comply with the provisions of the final order.

(xiv) The Certified Poultry Technician is intemperate in the use of stimulants, narcotics or other substances which impair the technician in their duties.

(b) *Appeal and request for a hearing.* An applicant or Certified Poultry Technician may request a hearing, in writing, within fifteen (15) days of receipt of notice of the proposed denial, suspension or revocation of their Certified Poultry Technician license or any proposed civil penalties to be assessed by the Department, as allowed under section 2383(b) of the Domestic Animal Law (3 Pa.C.S. § 2383(b)). The written request shall be sent to the Bureau of Animal Health and Diagnostic Services, Agriculture Building, 2301 North Cameron Street, Harrisburg, Pennsylvania 17110. The written request for a hearing must clearly set forth the basis of any appeal, and clearly identify the relevant issues or objections to be resolved. If you deny or challenge any averment in the Department's proposed order of denial, revocation or suspension, the appeal notice must identify that averment by number and describe the general basis for your denial or challenge. The scope of any subsequent administrative hearings or proceedings would be limited to those issues and objections set forth in the written appeal notice. No suspension, denial or revocation of licensure or civil penalty issued shall become effective until the time to appeal the proposed action has expired and the proposed order has thus become final or, where a hearing is requested, a hearing in the matter has been completed and a final order of the Department has been issued.

(c) *Revocation or suspension.* The Department's decision to deny, revoke or suspend a Certified Poultry Technician's licensure will be based on the gravity of the offense. The Department will consider such factors as the willfulness of the violation, previous violations and whether the person in question has continued to act as a Certified Poultry Technician after licensure was denied, suspended or revoked.

(i) *Suspended license.* A Certified Poultry Technician whose license has been suspended may not operate until that person has completed the required certification and licensure requirements or any final order issued by the Department, or both.

(ii) *Denied or Revoked license.* A Certified Poultry Technician whose certification or licensure has been denied or revoked may not operate until the Department has issued a final license. To obtain final certification and licensure, the person whose certification and license has been denied or revoked shall satisfy the full certification and licensing requirements established by this Order and shall have complied with any final order of the Department, including payment of any civil penalties and compliance with any requirements of such final order.

15. *Criminal and civil penalties.* The Department may impose those criminal and civil penalties specifically established at section 2383 of the Domestic Animal Law (3 Pa.C.S. § 2383) for any violation of the provisions of this Order.

16. *Civil remedy.* In addition to any other action or remedy sought, the Department may seek any civil remedy, as specifically established at section 2386 of the Domestic Animal Law (3 Pa.C.S. § 2386), for any violation of the provisions of this Order.

17. This Order shall not be construed as limiting the Department's authority to establish additional requirements for initial or continued certification of Certified Poultry Technicians or to limit any authority the Department possesses under the Domestic Animal Law, the CPT Act or any other Act or Law administered by the Department.

18. This Order rescinds, repeals and replaces the “General Quarantine Order; Certified Poultry Technicians” [49 Pa.B. 3088, Saturday, June 15, 2019]. This Order is effective upon the date of signature of the Secretary.

RUSSELL REDDING,
Secretary

[Pa.B. Doc. No. 25-423. Filed for public inspection March 28, 2025, 9:00 a.m.]

SECTION X: DANGEROUS TRANSMISSIBLE DISEASES OF POULTRY

Pennsylvania Dangerous Transmissible Diseases of Poultry (Diseases which may result in regulatory action)

- **Avian influenza**
- **Newcastle Disease**
- **Pullorum disease (*Salmonella pullorum*)**
- **Fowl typhoid (*S. gallinarum*)**
- **Avian mycoplasmosis (MG, MS)**
- **Duck viral enteritis**
- **Avian chlamydiosis (psittacosis, ornithosis)**

SECTION XI: OFFICIAL PDA LEG BANDS

Official Leg Band Requirements:

- Each blood sample tube for laboratory pullorum testing must be labeled with the band number of the bird from which the sample was collected or with a corresponding number on the submission form, so that the sample can be traced back to an individual bird.
- If testing a flock on site using the rapid pullorum test, all reactors must be banded, and either blood from the reactor is sent immediately to the laboratory for further testing, or the bird is euthanized and sent immediately to the laboratory for tissue culture.
- If testing birds for exhibition using the rapid pullorum test, band the birds before testing.
- All birds entering a show must be banded with an official leg band, even if exempt from pullorum testing.
- Leg band applicators are not provided by PDA.
- Records of leg band applications must be maintained for a minimum of two years by the CPT and must be made available to PDA upon request.

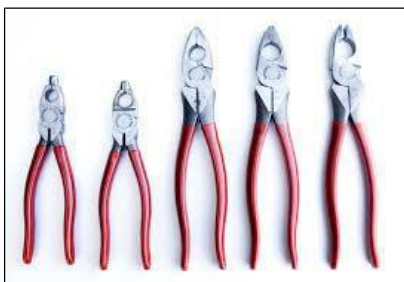
Official Leg Band Size Charts (Butt-End Style Leg Bands):

(Approximate Sizes) Butt-End Style Leg Band Size Chart



Band Size #	Inside Diameter	Recommendation
4	1/8"	Canaries, day old Quail
5	5/32"	Parakeet, 1-4 week old Quail, Doves, Cockatiels, Guinea keets
8	1/4"	Baby Chicken, Coturnix Quail, small Pigeon, adult Bobwhite Quail
10	5/16"	Pheasant hen, Tumblers, medium Pigeon, Homers
12	3/8"	Pheasant males, month old Chickens, large Pigeons, Chukar Partridge, Wood Duck
14	7/16"	Ringneck Pheasants, Mallards, Wild Ducks, Bantams, Feather Leg and Runt Pigeons
18	9/16"	Wild Ducks, Pheasants, Leghorn hens, Ancona, Silkie, Gamebirds
20	5/8"	Wild Turkey, Minorcas, Cross breeds, adult Guineas
22	11/16"	Plymouth Rock hens, Wyandottes, Rhode Island Red hens, Leghorn males, medium Ducks: Runners, Swedish, Harlequin
24	3/4"	Wild Geese, Plymouth Rock Males, Turkey hens, Geese, Orpingtons, larger ducks: Pekins, Rouens, Muscovy
28	7/8"	Canada Geese, Turkeys, Honkers

Each band size requires an appropriately sized applicator. Regular plier-type tools may also be effective in applying bands.



National Band & Tag Company
721 York St., PO Box 72430
Newport KY 41072-0430
USA 859-261-2035

Official Leg Band Distribution Sheet

[illegible]

SECTION XII: PA EXHIBIT BIRD RAPID PULLORUM PLATE TEST FORM

**PENNSYLVANIA DEPARTMENT OF AGRICULTURE
FLOCK TESTING REPORT FOR RAPID WHOLE BLOOD AGGLUTINATION PLATE
TEST (BIRD-SIDE TEST) FOR BIRDS NOT ENROLLED IN NPIP**

Name of flock owner: _____

Address of flock owner: _____

Location of flock (Premises owner/address): _____

Premise ID # of flock location: _____

Breed/strain/trade name of birds: _____

Number of birds in flock: _____

Age of birds: _____

Blood testing for Pullorum-typhoid (blood samples from all reactors on the rapid plate test, or the reactors, must be sent to a PADLS laboratory for further testing):

Number of males tested	Number of females tested	Number of reactors	Number of samples sent to laboratory	Laboratory results

Name of Certified Poultry Technician (please print): _____

Signature of Certified Poultry Technician: _____

Date: _____

A copy of this completed form must be sent to the PA Department of Agriculture BAHDS as soon as possible after testing has been completed. Keep a copy of this form and any laboratory results for your records. Remember that all exhibition birds tested for Pullorum using this test must be officially identified with a PDA leg band.

PA Department of
Agriculture
BAHDS - POULTRY
2301 N Cameron St
Harrisburg PA 17110
717-783-6897

PDA rev 12/12/22

SECTION XIII: AVIAN INFLUENZA FACTS

Avian Influenza Facts

Background: Avian influenza (AI) is a viral infection of birds, including wild birds, domestic poultry, pet birds, and birds in zoological collections. Low pathogenicity avian influenza (LPAI) infections are the most common infections identified in domestic poultry and may cause mild signs of disease, including respiratory signs, and decreased egg production in layer and breeder flocks. The morbidity and mortality in an infected flock is usually low unless there are secondary infections. Several AI viruses are highly pathogenic, causing severe systemic disease with multiple organ failure and high mortality. Highly pathogenic avian influenza (HPAI) is usually associated with H5 and H7 AI virus types.

HPAI viruses often cause severe, systemic disease with high mortality in chickens, turkeys, and other gallinaceous poultry. Waterfowl may also be affected. HPAI can cause high mortality in infected flocks, often as high as 100% within several days. There may be no clinical signs or gross lesions observed before death. In acute cases, lesions may include cyanosis and edema of the head, comb, wattle, and snood (turkey); ischemic necrosis of comb, wattles, or snood; edema and red discoloration of the legs and feet, hemorrhages on visceral organs and in muscles; and blood-tinged oral and nasal discharges. In severely affected birds, greenish diarrhea is common.

In 2022, a Eurasian H5 strain of highly pathogenic avian influenza (HPAI) was first detected in wild birds and domestic poultry in the United States. Infection in dairy cattle was first reported in March, 2024. For updates on this outbreak in poultry and other animals, refer to the USDA website at: [Detections of Highly Pathogenic Avian Influenza](#).

Diagnosis: Diagnosis can be confirmed through virus detection testing of material from poultry swabs or milk from lactating dairy cattle.

Treatment: There is no approved specific treatment available.

Prevention: The best method of preventing infection in a flock or dairy herd is to use the best biosecurity practices possible on the animal premises. Separation of dairy animals and poultry on a multi-species premises is very important.

Vaccination: Although vaccines exist, the use vaccination against AI in the United States requires a declaration of emergency and approval by the USDA Secretary.

Zoonosis: Although there is zoonotic potential, reports of human cases of AI are infrequent, and have been reported mostly where there is close contact between people and infected poultry or dairy cattle. Refer to the CDC website for more information.

Reporting: All suspected cases of avian influenza must be reported to the state veterinarian's office (717-772-2852, option 1 (24/7)).

Reference Sources: The Merck Veterinary Manual, David E. Swayne DVM, PhD, DACVP, DACPV, Southeast Poultry Research Laboratory, US National Poultry Research Center, Agricultural Research Service, US Department of Agriculture. USDA.

SECTION XIV: SUMMARY OF POULTRY PROGRAMS AND TESTING REQUIREMENTS FOR AVIAN INFLUENZA AND SALMONELLA PULLORUM IN PA

SUMMARY OF POULTRY PROGRAMS AND TESTING REQUIREMENTS FOR AVIAN INFLUENZA AND SALMONELLA PULLORUM IN PENNSYLVANIA

Table of Contents

The information provided here is only an overview of the requirements. Programs and testing requirements may change — please refer to the most current NPIP Provisions (9CFR), the PA Department of Agriculture, or individual exhibition managers for current requirements.

AI AGID testing of eggs of gallinaceous poultry may be allowed in some cases with prior permission.

Breeder & Commercial Flocks

- Section I: Layers
- Section II: Broilers
- Section III: Turkeys
- Section IV: Waterfowl, Hobbyist, and Exhibition-type Birds

Exhibition Only

- Section V: Birds for Exhibition/Shows

Game Birds

- Section VI: Game Birds

Section I: Layers

Layers - Avian Influenza

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. AI Clean 145 G	AI	Breeders, Primary flocks	On farm	Blood or T/OP swabs	30	30 birds tested negative initially, then 30 birds every 90 days or 30 birds during each 90 day period; AND 11 birds tested negative within 21 days prior to disposal/slaughter	≥ 4 months of age
NPIP U.S AI Clean 145 B	AI	Breeders, Multiplier flocks	On farm	Blood or T/OP swabs	30	30 birds tested negative initially, then 30 birds every 90 days or 30 birds during each 90 day period; AND 11 birds tested negative within 21 days prior to disposal/slaughter	≥ 4 months of age
NPIP U.S. H5/H7 AI Monitored (LPAI Voluntary Control Program) 146 B	AI	Commercial table-egg Layers	On farm	Blood or T/OP swabs	11	11 birds every 12 months AND 11 birds tested negative within 21 days prior to disposal/slaughter; AND pullets must be tested before placement in layer house	Should enroll in program if ≥ 75,000 layers on premises H5/H7
NPIP U.S. H5/H7 AI Monitored (LPAI Voluntary Control Program) 146 B	AI	Commercial table-egg Pullets	On farm	Blood or T/OP swabs	11	Test 11 pullets within 21 days of movement	Should enroll in program if ≥ 75,000 pullets total per year H5/H7

Layers - Salmonella pullorum

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. Pullorum-Typhoid Clean 145 G	Pullorum/Typhoid	Breeders; Primary Flock	On farm	Blood	300 or entire flock every 12 months	Every 12 months and 4 weeks after molt	≥4 months of age
NPIP U.S. Pullorum-Typhoid Clean 145 B	Pullorum/Typhoid	Breeders; Multiplier Flock (from P/T Clean primary breeder flock)	On farm	Blood	300 birds or entire flock every 12 months; at PDA discretion, may be exempt from annual test	Every 12 months and 4 weeks after molt	≥ 4 months of age

Section II: Broilers

Broilers – Avian Influenza

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. AI Clean 145 H	AI	Breeders; Primary flocks	On farm	Blood (pending- change proposed in 2024 to accept swabs)	30	30 birds tested negative initially, then 30 birds every 90 days or 30 birds during each 90 day period; AND up to 30 birds tested negative within 21 days prior to disposal/slaughter	≥ 4 months of age
NPIP U.S. AI Clean 145 C	AI	Breeders; Multiplier flocks	On farm	Blood (pending- change proposed in 2024 to accept swabs)	30	30 birds tested negative initially, then 15 birds every 90 days or 15 birds during each 90 day period; AND up to 30 birds tested negative within 21 days prior to disposal/slaughter	≥4 months of age
NPIP U.S. H5/H7 AI Monitored (LPAI Voluntary Control Program) 146C	AI	Commercial Broiler flocks - slaughter plants	On farm or at plant	Blood or T/OP swabs at plant; blood if sampled on farm. (pending- change proposed in 2024 to accept swabs for all testing)	11	11 birds tested negative within 21 days prior to slaughter or 11 birds per slaughter plant shift	FSIS-inspected slaughter plants processing 200,000 or more chickens/week. H5/H7

Broilers - Salmonella pullorum

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. Pullorum-Typhoid Clean 145 H	Pullorum/Typhoid	Breeders; Primary Flock	On farm	Blood	300 birds or entire flock every 12 months	Every 12 months and 4 weeks after molt	≥ 4 months of age
NPIP U.S. Pullorum-Typhoid Clean 145 C	Pullorum/Typhoid	Breeders; Multiplier Flock (from P/T Clean primary breeder flock)	On farm	Blood	300 birds or entire flock every 12 months; at PDA discretion, may be exempt from annual test	Every 12 months and 4 weeks after molt	≥ 4 months of age

Section III: Turkeys

Turkeys – Avian Influenza

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. AI Clean 145 D	AI	Breeders; Primary Flock	On farm	Blood or T/OP swabs	30	30 birds tested negative initially, then 30 birds every 90 days or 30 birds during each 90 day period; AND minimum 6 birds tested negative within 21 days prior to disposal/slaughter	≥ 4 months of age, prior to onset of egg production; may test after 12 weeks; H5/H7
NPIP U.S. AI Clean 145 D	AI	Breeders; Multiplier Flock	On farm	Blood	30	30 birds tested negative initially, then 30 birds every 90 days or 30 birds during each 90 day period; AND minimum 6 birds tested negative within 21 days prior to disposal/slaughter	≥ 4 months of age, prior to onset of egg production; may test after 12 weeks; H5/H7
NPIP U.S. H5/H7 AI Monitored (LPAI Voluntary Control Program) 146 D	AI	Commercial Turkey Flocks	On farm or at slaughter	Blood or T/OP swabs	Minimum of 6 (Most participants test 11 birds)	Minimum 6 birds per flock within 21 days prior to slaughter or equivalent number tested at plant	FSIS inspected slaughter plants processing 2 million or more turkeys/year. H5/H7. It is recommended that samples be collected from flocks over 10 weeks of age with respiratory signs; depression; or decreases in food or water intake.

Turkeys - *Salmonella pullorum*

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. Pullorum-Typhoid Clean 145 D	Pullorum/Typhoid	Breeders; Primary Flock	On farm	Blood (Rapid whole blood plate test is NOT APPROVED for turkeys)	300 or entire flock every 12 months	Every 12 months and 4 weeks after molt	≥ 4 months, may test after 12 weeks of age
NPIP U.S. Pullorum-Typhoid Clean 145 D	Pullorum/Typhoid	Breeders; Multiplier Flock (from P/T Clean primary breeder flock)	On farm	Blood (Rapid whole blood plate test is NOT APPROVED for turkeys)	300 birds or entire flock every 12 months; at PDA discretion, may be exempt from annual test	Every 12 months and 4 weeks after molt	≥ 4 months, may test after 12 weeks of age

Section IV: Waterfowl, Hobbyist, and Exhibition-type Poultry

Waterfowl, Hobbyist, and Exhibition-type Poultry – Avian Influenza

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. H5/H7 AI Clean Hobbyist, Exhibition poultry, Raised for release waterfowl breeding flocks/products 145 E	AI	Hobbyist & Exhibition- type Poultry, Raised-for- Release Waterfowl Breeding Flocks/prod ucts	On farm	Gallinaceous birds-blood or T/OP swabs Waterfowl- Cloacal swabs	30	30 birds tested negative initially, then 30 birds every 180 days or 30 birds during each 180 day period; AND 30 birds tested negative within 21 days prior to disposal/slaughter	≥ 4 months, or sexual maturity (game birds); H5/H7 Note: Mixed-species hatcheries are included under this NPIP subpart
NPIP U.S. H5/H7 AI Clean Primary meat-type waterfowl flocks/products 145 I	AI	Primary Meat-type Waterfowl Breeding Flocks	On farm	Cloacal swabs	30	30 birds tested negative initially, then 30 birds every 90 days or 30 birds during each 90 day period AND 11 birds tested negative within 21 days prior to disposal/slaughter	≥ 4 months H5/H7
NPIP U.S. H5/H7 AI Clean Multiplier meat-type waterfowl flocks/products 145 I	AI	Multiplier Meat- type Waterfowl Breeding Flocks	On farm	Cloacal swabs	30	30 birds tested negative initially, then 30 birds every 180 days or 30 birds during each 180 day period AND 11 birds tested negative within 21 days prior to disposal/slaughter	≥ 4 months H5/H7
NPIP H5/H7 LPAI Monitoring Program – 146 E	AI	Waterfowl slaughter plants	At slaughter or on farm	Waterfowl - Cloacal swabs	11	Per shift or on farm within 21 days prior to slaughter	Plant processes 50,000 or more birds per year. H5/H7
NPIP H5/H7 LPAI Monitoring Program 146 E	AI	Waterfowl flocks producing eggs for human consumption	On farm	Waterfowl - Cloacal swabs	11	11 birds tested within 30 days of disposal/slaughter or within a 12- month period	Waterfowl premises with ≥ 25,000 birds. H5/H7

Waterfowl, Hobbyist, and Exhibition-type Poultry - Salmonella pullorum/typhoid

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. Pullorum-Typhoid Clean Hobbyist, Exhibition-type poultry, Raised for release waterfowl breeding flocks/products 145 E	Pullorum/Typhoid	Hobbyist, Exhibition-type, Raised for release waterfowl breeding flocks/products Breeders	On farm	Blood	300 birds or entire flock every 12 months.	Every 12 months and 4 weeks after molt	≥ 4 months of age, or sexual maturity (game birds)
NPIP U.S. Pullorum-Typhoid Clean Meat type waterfowl breeding flocks/products 145 I	Pullorum/Typhoid	Meat type waterfowl Breeders	On farm	Blood	300 birds or entire flock every 12 months.	Every 12 months and 4 weeks after molt	≥ 4 months of age

Birds for PA Exhibition/Show

Section V: Exhibition Birds (All birds entering a show must be banded with official PDA leg bands)

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
AI testing for birds going to PA exhibition/show	AI	Exhibition birds for show: See current PDA Quarantine Order for Exhibition (market birds & non-poultry allowed)	On farm	Gallinaceous birds: Blood or T/OP swabs Waterfowl: cloacal swabs are required	30 birds, representative of the entire flock, must be tested negative on an official test within the 14 days prior to entry into the exhibition	Within 14 days prior to entry into the exhibition	Official PDA leg band if showing Non-poultry definition & other requirements: See Current PA Avian Exhibition Quarantine Order
Pullorum testing for PA birds going to PA exhibition	Pullorum/typhoid	Exhibition birds for show (waterfowl and market poultry exempt)	On farm	Blood	All poultry except waterfowl & market birds going to exhibition must come from NPIP P/T Clean Flock (flock must be enrolled in NPIP), OR a PA Pullorum Equivalent Program Flock, OR must be tested within 90 days prior to entry. If too young to test, must come from a tested flock.	Within 90 days prior to entry into the exhibition	Official PDA leg band
PA Pullorum Equivalent Program	Pullorum/typhoid	Exhibition poultry, except waterfowl	On farm	Blood	300 or entire flock if less than 300 birds	Every 12 months	Official PDA leg bands. Must enroll with PDA. Annual inspection.

Upland Game Birds

Section VI: NPIP Upland Game Birds

Program	Disease	Type of Bird	Location	Type of Sample	# of Birds Tested	Frequency of Testing	Comment
NPIP U.S. H5/H7 AI Clean 145 J Egg/Meat-type Game Bird and Raised for release game bird breeding flocks/products	AI	Hatcheries, Breeders, Growers, Dealers, Raised for Release	On farm	Blood or T/OP swabs	30	30 birds every 90 days or 30 birds during each 90 day period	≥ 4 months or sexual maturity H5/H7 For participants with non- breeding flocks retained for raised-for-release or other purposes on the same premises as a breeding flock, these birds must be included in AI testing every 90 days. (R for R birds enrolled in 145 J)
NPIP U.S. H5/H7 Pullorum Clean 145 J Egg/Meat-type Game Bird and Raised for release game bird breeding flocks)	Pullorum/typhoid	Hatcheries, Breeders, Growers, Dealers, Raised for Release	On farm	Blood	300 birds or entire flock every 12 months.	Every 12 months and 4 weeks after molt	≥ 4 months or sexual maturity
NPIP H5/H7 LPAI Monitoring Program 146E	AI	Game Bird slaughter plants	At slaughter or on farm	Blood or T/OP swabs	11	Per shift or on farm within 21 days prior to slaughter	Plant processes 50,000 or more birds per year H5/H7
NPIP H5/H7 LPAI Monitoring Program 146E	AI	Game Birds producing eggs for human consumption	On farm	Blood or T/OP swabs	11	11 birds tested within 30 days of disposal/slaughter or within a 12- month period	Egg-type game bird premises with ≥ 25,000 birds H5/H7